

(FOR RESEARCH USE ONLY. DO NOT USE IT IN CLINICAL DIAGNOSIS !)

**Catalog No: E-BC-F001**

**Specification: 48T(32 samples)/96T(80 samples)/500Assays(484 samples)**

**Measuring instrument: Fluorescence Microplate Reader  
(Ex/Em=535 nm/587 nm)**

**Detection range: 0.02-10  $\mu$ mol/L**

## **Elabscience<sup>®</sup>Hydrogen Peroxide (H<sub>2</sub>O<sub>2</sub>) Fluorometric Assay Kit**

This manual must be read attentively and completely before using this product.

If you have any problem, please contact our Technical Service Center for help:

Toll-free: 1-888-852-8623

Tell: 1-832-243-6086

Fax: 1-832-243-6017

Email: [techsupport@elabscience.com](mailto:techsupport@elabscience.com)

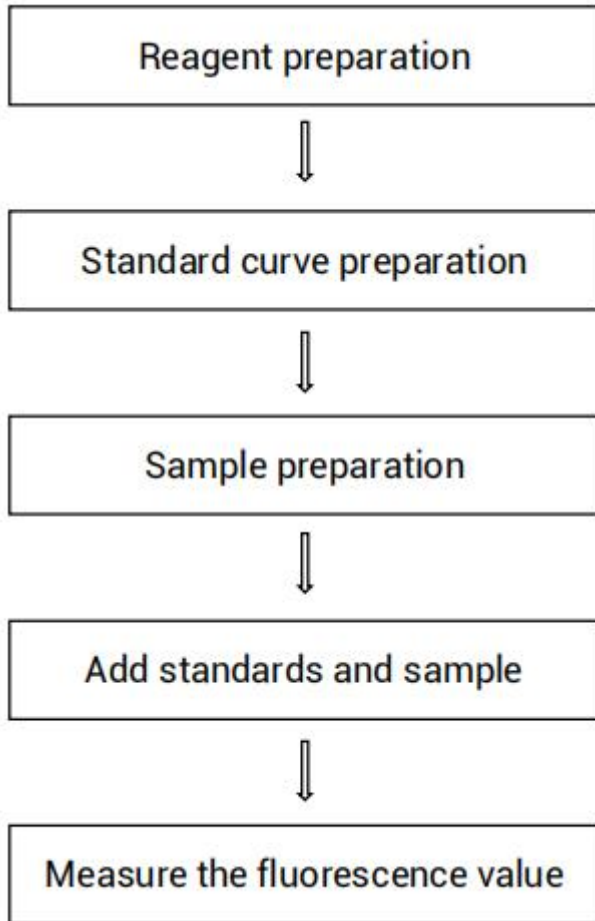
Website: [www.elabscience.com](http://www.elabscience.com)

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

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## Assay summary



## Intended use

This kit can be used to measure the H<sub>2</sub>O<sub>2</sub> content in serum, plasma, tissue and cells samples.

## Detection principle

In the presence of peroxidase, hydrogen peroxide reacts with the fluorescent probe, and the fluorescence intensity at the excitation wavelength of 535 nm and the emission wavelength of 587 nm is proportional to the hydrogen peroxide concentration.

## Kit components & storage

Item	Component	Size 1(48 T)	Size 2(96 T)	Size 3 (500 Assays)	Storage
Reagent 1	Buffer Solution	30 mL × 1 vial	60 mL × 1 vial	60 mL × 5 vials	-20°C, 12 months
Reagent 2	Substrate	0.06 mL × 1 vial	0.12 mL × 1 vial	0.12 mL × 5 vials	-20°C, 12 months shading light
Reagent 3	Enzyme Reagent	Powder × 1 vial	Powder × 1 vial	Powder × 5 vials	-20°C, 12 months shading light
Reagent 4	1 mol/L H <sub>2</sub> O <sub>2</sub> Standard Solution	0.1 mL × 1 vial	0.1 mL × 1 vial	0.1 mL × 5 vials	-20°C, 12 months
Reagent 5	Protein Precipitator	10 mL × 1 vial	20 mL × 1 vial	20 mL × 5 vials	2-8°C, 12 months
Reagent 6	Alkali Reagent	3 mL × 1 vial	6 mL × 1 vial	6 mL × 5 vials	2-8°C, 12 months
	pH Test Strips	1 bag		5 bags	
	Black Microplate	96 wells			No requirement
	Plate Sealer	2 pieces			
	Sample Layout Sheet	1 piece			

Note: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other. For a small volume of reagents, please centrifuge before use, so as not to obtain sufficient amount of reagents.

## **Materials prepared by users**

### **Instruments:**

Fluorescence microplate reader (Ex/Em=535 nm/587 nm), Micropipettor, Incubator, Vortex mixer, Centrifuge

## **Reagent preparation**

- ① Equilibrate all the reagents to room temperature before use.
- ② The preparation of enzyme application solution:  
Dissolve one vial of enzyme reagent with 120  $\mu\text{L}$  of buffer solution. Mix well to dissolve. Store at  $-20^{\circ}\text{C}$  for 1 month protected from light.
- ③ The preparation of working solution:  
Before testing, please prepare sufficient working solution according to the test wells. For example, prepare 250  $\mu\text{L}$  of working solution (mix well 240  $\mu\text{L}$  of buffer solution, 5  $\mu\text{L}$  of substrate and 5  $\mu\text{L}$  of enzyme application solution). The working solution should be prepared on spot.
- ④ The preparation of 10 mmol/L  $\text{H}_2\text{O}_2$  solution:  
Dilute 10  $\mu\text{L}$  of 1 mol/L  $\text{H}_2\text{O}_2$  solution with 990  $\mu\text{L}$  of buffer solution.  
Mix well to dissolve.
- ⑤ The preparation of 100  $\mu\text{mol/L}$   $\text{H}_2\text{O}_2$  solution:  
Dilute 10  $\mu\text{L}$  of 10 mmol/L  $\text{H}_2\text{O}_2$  solution with 990  $\mu\text{L}$  of buffer solution.  
Mix well to dissolve.

- ⑥ The preparation of 10  $\mu\text{mol/L}$   $\text{H}_2\text{O}_2$  solution:  
Dilute 70  $\mu\text{L}$  of 100  $\mu\text{mol/L}$   $\text{H}_2\text{O}_2$  solution with 630  $\mu\text{L}$  of buffer solution.  
Mix well to dissolve.
- ⑦ The preparation of standard curve:  
Dilute 10  $\mu\text{mol/L}$   $\text{H}_2\text{O}_2$  solution with buffer solution to a serial concentration. The recommended dilution gradient is as follows: 0, 0.5, 1, 2, 4, 6, 8, 10  $\mu\text{mol/L}$ . Reference is as follows:

Item	①	②	③	④	⑤	⑥	⑦	⑧
<b>Concentration (<math>\mu\text{mol/L}</math>)</b>	<b>0</b>	<b>0.5</b>	<b>1</b>	<b>2</b>	<b>4</b>	<b>6</b>	<b>8</b>	<b>10</b>
<b>10 <math>\mu\text{mol/L}</math> standard (<math>\mu\text{L}</math>)</b>	0	10	20	40	80	120	160	200
<b>Buffer solution (<math>\mu\text{L}</math>)</b>	200	190	180	160	120	80	40	0

## Sample preparation

### ① Sample preparation

**Serum, plasma and other liquid sample:** detect directly. If not detected on the same day, the serum or plasma can be stored at  $-80^\circ\text{C}$  for a month.

### **Tissue sample:**

- ① Harvest the amount of tissue needed for each assay (initial recommendation 20 mg).
- ② Wash tissue in cold PBS (0.01 M, pH 7.4).
- ③ Homogenize 20 mg tissue in 180  $\mu\text{L}$  buffer solution with a dounce homogenizer at  $4^\circ\text{C}$ .
- ④ Centrifuge at  $10000\times g$  for 10 min to remove insoluble material. Collect supernatant and keep it on ice for detection.
- ⑤ Meanwhile, determine the protein concentration of supernatant (E-BC-K318-M).

### Cell (adherent or suspension) samples:

- ① Harvest the number of cells needed for each assay (initial recommendation  $1 \times 10^6$  cells).
- ② Wash cells with PBS (0.01 M, pH 7.4).
- ③ Homogenize  $1 \times 10^6$  cells in 200  $\mu$ L buffer solution with a ultrasonic cell disruptor at 4°C.
- ④ Centrifuge at  $12000 \times g$  for 10 min to remove insoluble material. Collect supernatant and keep it on ice for detection.
- ⑤ Meanwhile, determine the protein concentration of supernatant (E-BC-K318-M).

### ② Dilution of sample

The recommended dilution factor for different samples is as follows (for reference only):

Sample type	Dilution factor
Human serum	1-3
Mouse serum	1-4
Mouse plasma	1-2
Porcine serum	1-3
Rat serum	1-4
HepG2 culture supernatant	1
10% Mouse liver tissue homogenate	1-3
10% Mouse brain tissue homogenate	1-3
10% Rat lung tissue homogenate	1-3
HepG2 cells homogenate (the concentration of protein is 6.44 gprot/L)	1-2

Note: The diluent is buffer solution. For the dilution of other sample types, please do pretest to confirm the dilution factor

## The key points of the assay

- ① Avoid repeated freezing and thawing of substrate, it is recommended to aliquot the substrate into smaller quantities and store at  $-20^{\circ}\text{C}$ .
- ② Because  $\text{H}_2\text{O}_2$  is very unstable, prepare the  $\text{H}_2\text{O}_2$  standard solution freshly.
- ③ The prepared working solution must be stored with shading light.
- ④ The pH of pretreated sample should be 6.5-8.
- ⑤ Prevent the formulation of bubbles when the supernatant is transferred into the microplate.

## Operating steps

### The pretreatment of sample

For each well, add  $150\ \mu\text{L}$  of sample and  $150\ \mu\text{L}$  of protein precipitator, mix well.

Centrifuge at  $13000\times g$  for 10 min, then take  $V_1$  (eg,  $V_1=0.2\ \text{mL}$ ) of the supernatant, add  $V_2$  (eg,  $V_2=0.04\ \text{mL}$ ) of alkali reagent to adjust the pH to 6.5-8, which is the sample to be tested.

### The measurement of samples

- ① Standard tube: add  $50\ \mu\text{L}$  of standard solution with different concentrations to the wells.  
Sample tube: add  $50\ \mu\text{L}$  of sample to the wells.
- ② Add  $50\ \mu\text{L}$  of working solution to each well.
- ③ Mix fully with microplate reader for 10 s and incubate the plate at room temperature for 10 min with shading light.
- ④ Measure the fluorescence intensity at the excitation wavelength of 535 nm and the emission wavelength of 587 nm.

## Calculation

### The standard curve:

1. Average the duplicate reading for each standard.
2. Subtract the mean fluorescence value of the blank (Standard #①) from all standard readings. This is the absolved fluorescence value.
3. Plot the standard curve by using absolved fluorescence value of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve ( $y = ax + b$ ) with graph software (or EXCEL).

### The sample:

#### 1. Serum, plasma and other liquid samples:

$$\text{H}_2\text{O}_2 \text{ content } (\mu\text{mol/L}) = (\Delta F - b) \div a \times 2 \times \left( \frac{V_1 + V_2}{V_1} \right) \times f$$

#### 2. Tissue and cell samples:

$$\text{H}_2\text{O}_2 \text{ content } (\mu\text{mol/gprot}) = (\Delta F - b) \div a \times 2 \times \left( \frac{V_1 + V_2}{V_1} \right) \times f \div C_{\text{pr}}$$

### [Note]

$\Delta F$ : Absolute fluorescence intensity of sample ( $F_{\text{Sample}} - F_{\text{Blank}}$ )

f: Dilution factor of sample before tested.

2: Dilution factor of sample in pretreatment step ( $V_{\text{sample}}:V_{\text{protein precipitator}} = 1:1$ ).

$V_1$ : The volume of supernatant in pretreatment step (mL).

$V_2$ : The volume of alkali reagent in pretreatment step (mL).

$C_{\text{pr}}$ : Concentration of protein in sample (gprot/L).

## Appendix I Performance Characteristics

### 1. Parameter:

#### Intra-assay Precision

Three human serum samples were assayed in replicates of 20 to determine precision within an assay. (CV = Coefficient of Variation)

Parameters	Sample 1	Sample 2	Sample 3
Mean ( $\mu\text{mol/L}$ )	1.20	3.50	8.00
%CV	1.3	1.0	1.0

#### Inter-assay Precision

Three human serum samples were assayed 20 times in duplicate by three operators to determine precision between assays.

Parameters	Sample 1	Sample 2	Sample 3
Mean ( $\mu\text{mol/L}$ )	1.20	3.50	8.00
%CV	3.4	4.1	3.3

#### Recovery

Take three samples of high concentration, middle concentration and low concentration to test the samples of each concentration for 6 times parallelly to get the average recovery rate of 100%.

	Standard 1	Standard 2	Standard 3
Expected Conc. ( $\mu\text{mol/L}$ )	0.9	4.2	8.5
Observed Conc. ( $\mu\text{mol/L}$ )	0.9	4.2	8.8
Recovery rate (%)	96	100	104

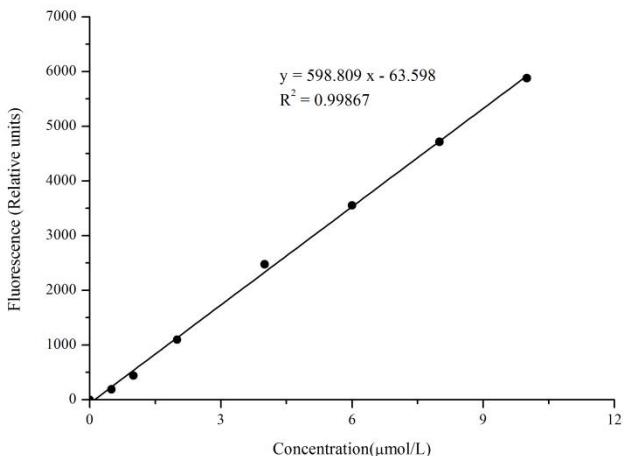
#### Sensitivity

The analytical sensitivity of the assay is 0.02  $\mu\text{mol/L}$ . This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 20 times, and calculating the corresponding concentration.

## 2. Standard curve:

As the fluorescence value of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique or temperature effects), so the standard curve and data are provided as below for reference only:

Concentration ( $\mu\text{mol/L}$ )	0	0.5	1	2	4	6	8	10
Fluorescence value	119	311	564	1226	2601	3691	4873	5998
	122	309	557	1210	2589	3658	4800	6002
Average fluorescence value	120	310	561	1218	2595	3674	4836	6000
Absoluted fluorescence value	0	190	441	1098	2475	3554	4716	5880



## Appendix Π Example Analysis

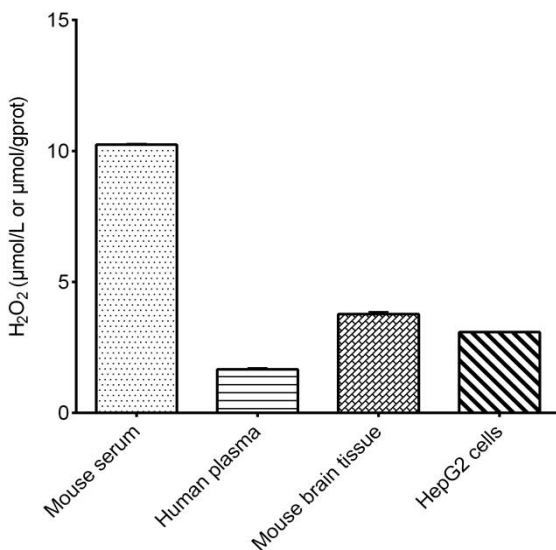
### Example analysis :

For 10% mouse brain tissue homogenate, take 50  $\mu\text{L}$  and carry the assay according to the operation steps. The results are as follows:

Standard curve:  $y = 556.7x + 11.559$ , the average fluorescence value of the sample is 2452, the average fluorescence value of the blank is 80, the concentration of protein in sample is 5.4 gprot/L, and the calculation result is:

$$\begin{aligned} \text{H}_2\text{O}_2 \text{ content} \\ (\mu\text{mol/gprot}) &= (2452 - 80 - 11.559) \div 556.7 \times 2 \times ((0.2 + 0.04) \div 0.2) \times 2 \div 5.4 \\ &= 3.77 \mu\text{mol/gprot} \end{aligned}$$

Detect mouse serum, human plasma, 10% mouse brain tissue homogenate (the concentration of protein is 5.4 gprot/L, dilute for 2 times) and HepG2 cells (the concentration of protein is 6.44 gprot/L) according to the protocol, the result is as follows:



## Statement

1. This assay kit is for Research Use Only. We will not response for any arising problems or legal responsibilities causing by using the kit for clinical diagnosis or other purpose.
2. Please read the instructions carefully and adjust the instruments before the experiments. Please follow the instructions strictly during the experiments.
3. Protection methods must be taken by wearing lab coat and latex gloves.
4. If the concentration of substance is not within the detection range exactly, an extra dilution or concentration should be taken for the sample.
5. It is recommended to take a pre-test if your sample is not listed in the instruction book.
6. The experimental results are closely related to the situation of reagents, operations, environment and so on. Elabscience will guarantee the quality of the kits only, and NOT be responsible for the sample consumption caused by using the assay kits. It is better to calculate the possible usage of sample and reserve sufficient samples before use.





