

## **CIM (Cimaterol) ELISA Kit**

Catalog No: E-FS-E157

96T/96T\*3

This manual must be read attentively and completely before using this product.

If you have any problems, please contact our Technical Service Center for help.

Toll-free: 1-888-852-8623 Tel: 1-832-243-6086 Fax: 1-832-243-6017

Email: [techsupport@elabscience.com](mailto:techsupport@elabscience.com)

Website: [www.elabscience.com](http://www.elabscience.com)

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

## Test principle

This kit uses Competitive-ELISA as the method for the quantitative detection. It can detect Cimaterol (CIM) in samples, such as muscle, feed, etc. This kit is composed of ELISA Microtiter plate, HRP conjugate, antibody working solution, standard and other supplementary reagents. The microtiter plate in this kit has been pre-coated with coupled antigen. During the reaction, CIM in the samples or standard competes with coupled antigen on the solid phase supporter for sites of anti-CIM antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each microtiter plate well, and substrate reagent is added for color development. There is a negative correlation between the OD value of samples and the concentration of CIM. The concentration of CIM in the samples can be calculated by comparing the OD of the samples to the standard curve.

## Technical indicator

**Reaction mode**(Incubation time and temperature) : 25℃; 30 min, 15 min

**Detection limit:** Urine (Swine) ---1.5 ppb; Urine (Bovine) ---3 ppb; Muscle (method 1) ---2 ppb; Muscle (method 2) ---0.5 ppb; Serum---2 ppb

**Cross-reactivity:** Cimaterol---100%; Terbutaline---4%; Clenbuterol---40%; Salbutamol--8%;  
Brombuterol---50%; Tulobuterol ---15%; Clorprenaline—5%;  
Fenoterol, Penbutolol ---<0.1%

**Sample recovery rate:** 90% ± 30%.

## Kits components

Item	Specifications
ELISA Microtiter plate	96 wells
Standard Liquid	1.5 mL each (ppb=ng/mL=ng/g) (0 ppb, 0.2 ppb, 0.6 ppb, 1.8 ppb, 5.4 ppb, 16.2 ppb)
HRP Conjugate	6 mL
Antibody Working Solution	6 mL
Substrate Reagent A	7 mL
Substrate Reagent B	7 mL
Stop Solution	7 mL
20×Concentrated Wash Buffer	25 mL
Reconstitution Buffer	60 mL
Serum Diluent	60 mL
Plate Sealer	3 pieces
Sealed Bag	1 piece
Manual	1 copy

Note: All reagent bottle caps must be tightened to prevent evaporation and microbial pollution.

**Other materials required but not supplied**

**Instruments:** Microplate reader, Printer, Homogenizer, Nitrogen evaporators, Water bath, Vortex mixer, Centrifuge, Graduated pipette, Balance (sensitivity 0.01 g).

**Micropipette:** Single channel (20-200  $\mu\text{L}$ , 100-1000  $\mu\text{L}$ ), Multichannel (30-300  $\mu\text{L}$ ).

**Reagents:** NaOH, Ethyl acetate, Acetonitrile, N-hexane, Trichloroacetic acid

**Experimental preparation**

Restore all reagents and samples to room temperature before use.

Open the microplate reader in advance, preheat the instrument, and set the testing parameters.

**1. Sample pretreatment Notice:**

Experimental apparatus should be clean and the pipette should be disposable to avoid cross-contamination during the experiment.

**2. Solution preparation**

*Please prepare solution according to the number of samples. Don't use up all components in the kit at once!*

Solution 1: 0.5% Trichloroacetic acid Solution (*for pork, beef, mutton sample*)

Dissolve 1 g of **Trichloroacetic acid** to 200 mL with deionized water, mix fully.

Solution 2: 0.5 M NaOH Solution (*for pork, beef, mutton sample*)

Dissolve 2 g of **NaOH** to 100 mL with deionized water, mix fully.

Solution 3: Wash Buffer

Dilute **20×Concentrated Wash Buffer** with deionized water, mix fully.

(20×Concentrated Wash Buffer (V): Deionized water (V) = 1:19).

## Notes

1. The overall OD value will be lower when reagents have not been brought to room temperature before use or room temperature is below 25°C.
2. If the wells turn dry during the washing procedure, it will lead to bad linear standard curve and poor repeatability. Operate the next step immediately after wash.
3. Mix thoroughly and wash the plate completely. The consistency of wash procedure can strongly affect the reproducibility of this ELISA kit.
4. ELISA Microplate should be covered by plate sealer. Avoid the kit to strong light.
5. **Each reagent is optimized for use in the E-FS-E157. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other E-FS-E157 with different lot numbers.**
6. Substrate Reagent should be abandoned if it turns blue color. When OD value of standard (concentration: 0) < 0.8 unit (A450nm < 0.8), it indicates the reagent may be deteriorated.
7. Stop solution is caustic, avoid contact with skin and eyes.
8. As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.
9. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
10. **For mentioned sample fast and efficient extraction methods are included in the kit description. Please consult technical support for the applicability if other sample need to be tested.**
11. The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS, etc. can be used for quantitative confirmation.

## Storage and expiry date

Store the kit at 2-8°C. Do not freeze any test kit components.

Return any unused microwells to their original foil bag and reseal them together with the desiccant provided and further store at 2-8°C.

**Expiry date:** expiration date is on the packing box.

### 3. Sample pretreatment procedure

#### 3.1 Pretreatment of swine urine sample:

Take 50  $\mu\text{L}$  of clear urine sample for analysis directly (if the urine sample is turbid, it should be filtered or centrifuged at 4000 r/min for 5 min until the urine sample become clear). Samples temporarily not used should be frozen.

**Note: Sample dilution factor: 1, detection limit: 1.5 ppb.**

#### 3.2 Pretreatment of muscle (pork, beef, mutton) (method 1):

- (1) Remove fat from sample, homogenize the sample with homogenizer.
- (2) Weigh  $1 \pm 0.01$  g of crushed homogenate muscle sample, add 3 mL of **0.5% Trichloroacetic acid Solution** (Solution 1). Vortex fully for 2 min, centrifuge at 4000 r/min for 5 min.
- (3) Take 1 mL of the middle solution to another centrifuge tube, add 40  $\mu\text{L}$  of **0.5 M NaOH Solution** (Solution 2). Vortex for 2 min, centrifuge at 4000 r/min for 5 min.
- (4) Take 50  $\mu\text{L}$  of the supernatant for analysis.

**Note: Sample dilution factor: 4, detection limit: 2 ppb.**

#### 3.3 Pretreatment of muscle (livestock) (method 2):

- (1) Remove fat from sample, homogenize the sample with homogenizer.
- (2) Weigh  $3 \pm 0.05$  g of crushed homogenate muscle sample, add 8 mL of **Acetonitrile** and 1 mL of **Ethyl acetate**. Vortex fully for 2 min, centrifuge at 4000 r/min for 10 min.
- (3) Take 4 mL of the supernatant to another centrifuge tube, dry at  $60^\circ\text{C}$  with nitrogen evaporators or water bath.
- (4) Dissolve the residue with 1 mL of **N-hexane**, add 1 mL of **Reconstitution Buffer**, and mix fully for 30s. Centrifuge at 4000 r/min for 5 min at room temperature.
- (5) Discard the upper organic phase and middle solution, take 50  $\mu\text{L}$  of the lower water layer for analysis.

**Note: Sample dilution factor: 1, detection limit: 0.5 ppb.**

#### 3.4 Pretreatment of serum (Bovine) sample:

- (1) Take 200  $\mu\text{L}$  of serum to centrifuge tube. Add 600  $\mu\text{L}$  of **Serum Diluent** and vortex fully for 30s.
- (2) Incubation at  $80^\circ\text{C}$  for 5 min with water bath.
- (3) Take 50  $\mu\text{L}$  for analysis after the sample returns to room temperature ( $25^\circ\text{C}$ ).

**Note: Sample dilution factor: 4, detection limit: 2 ppb**

#### 3.5 Pretreatment of urine (Bovine) sample:

- (4) Take 100  $\mu\text{L}$  of urine to centrifuge tube. Add 200  $\mu\text{L}$  of **Deionized water** and vortex fully for 10s.
- (5) Take 50  $\mu\text{L}$  for analysis after the sample returns to room temperature ( $25^\circ\text{C}$ ).

**Note: Sample dilution factor: 3, detection limit: 2 ppb**

**Assay procedure**

Restore all reagents and samples to room temperature (25°C) before use. All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming. The unused ELISA Microtiter plate should be sealed as soon as possible and stored at 2-8°C.

1. **Number:** number the sample and standard in order (multiple well), and keep a record of standard wells and sample wells. **Standard and Samples need test in duplicate.**
2. **Add Sample:** add 50 µL of **Standard or Sample** per well, then add 50 µL **HRP Conjugate** to each well. Add 50 µL of **Antibody Working Solution**, cover the plate with plate sealer. Oscillate for 5s gently to mix thoroughly, incubation at 25°C for 30 min in shading light
3. **Wash:** uncover the sealer carefully, remove the liquid in each well. Immediately add 260µL of **Wash Buffer** (Solution 3) to each well and wash. Repeat wash procedure for 5 times, 30s intervals/time. Invert the plate and pat it against thick clean absorbent paper (If bubbles exist in the wells, clean tips can be used to prick them).
4. **Color Development:** add 50 µL of **Substrate Reagent A** to each well, and then add 50 µL of **Substrate Reagent B**. Gently oscillate for 5s to mix thoroughly. Incubate at 25°C for 15 min in shading light (The reaction time can be extended according to the actual color change).
5. **Stop Reaction:** add 50 µL of **Stop Solution** to each well, gently oscillate and mix fully to stop the reaction.
6. **OD Measurement:** determine the optical density (OD value) of each well at 450 nm (reference wavelength 630 nm) with a microplate reader. This step should be finished in 5 min after stop reaction.

## Result analysis

### 1. Absorbance% = $A/A_0 \times 100\%$

A: Average absorbance of standard solution or sample

$A_0$ : Average absorbance of 0 ppb Standard solution

### 2. Drawing and calculation of standard curve

Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add the average absorbance value of sample to standard curve to get corresponding concentration. **If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.**

For this kit, it is more convenient to use professional analysis form for accurate and fast analysis on many samples.

### Cimaterol (E-FS-E157) Standard Curve

