

(FOR RESEARCH USE ONLY. DO NOT USE IT IN CLINICAL DIAGNOSIS !)

Catalog No: E-BC-K071-M

Specification: 48T(31 samples)/96T(79 samples)/500Assays(483 samples)

Measuring instrument: Microplate reader (510-530 nm)

Detection range: 0.31-50 mg/L

Elabsience® Total Iron Binding Capacity (TIBC)

Colorimetric Assay Kit

This manual must be read attentively and completely before using this product.

If you have any problem, please contact our Technical Service Center for help:

Toll-free: 1-888-852-8623

Tell: 1-832-243-6086

Fax: 1-832-243-6017

Email: techsupport@elabsience.com

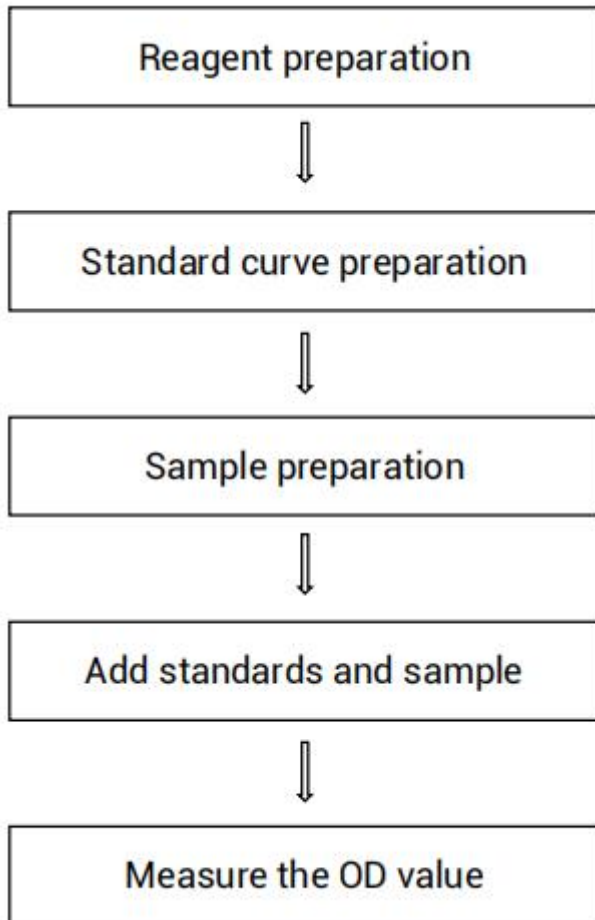
Website: www.elabsience.com

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

Table of contents

Assay summary	3
Intended use	4
Detection principle	4
Kit components & storage	5
Materials prepared by users	5
Reagent preparation	6
Sample preparation	7
The key points of the assay	7
Operating steps	8
Calculation	9
Appendix I Performance Characteristics	10
Appendix II Example Analysis	12
Statement	13

Assay summary



Intended use

This kit can be used to measure the total iron binding capacity (TIBC) content in serum samples.

Detection principle

The excess iron is added to the serum to bind all the ferritin in the serum, and the excess iron is adsorbed by adding the iron adsorbent. The iron bind with the ferritin is separated from the protein by the action of acid solution and reductant. Fe^{3+} in serum is reduced to Fe^{2+} , Fe^{2+} binds with bipyridine to form pink complex. In a certain range, the amount of TIBC is positively correlated with the depth of color. The iron content measured is, minus serum iron value, which is called unsaturated iron binding force. Total iron binding capacity minus serum iron value is unsaturated iron binding capacity (UIBC).

Kit components & storage

Item	Component	Size 1 (48 T)	Size 2 (96 T)	Size 3(Size 3) (500Assays)	Storage
Reagent 1	100 mg/L Iron Standard Stock Solution	2 mL × 1 vial	2 mL × 1 vial	2 mL×5 vials	2-8℃, 12 months
Reagent 2	Chromogenic Agent A	Powder × 1 vial	Powder × 2 vials	Powder × 10 vials	2-8℃, 12 months shading light
Reagent 3	Chromogenic Agent B	Powder × 1 vial	Powder × 2 vials	Powder × 10 vials	2-8℃, 12 months shading light
Reagent 4	Chromogenic Agent C	15 mL × 1 vial	15 mL × 2 vials	30 mL × 5 vials	2-8℃, 12 months
Reagent 5	Iron Absorbent	Powder × 31 vials	Powder × 79 vials	Powder × 483vials	2-8℃, 12 months
	Microplate	48 wells	96 wells	/	No requirement
	Plate Sealer	2 pieces		/	
	Sample Layout Sheet	1 piece			

Note: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other. For a small volume of reagents, please centrifuge before use, so as not to obtain sufficient amount of reagents.

Materials prepared by users

Instruments:

Microplate reader (510-530 nm, optimum wavelength: 520 nm),

Micropipettor, Water bath, Incubator, Vortex mixer, Centrifuge

Reagents:

Double distilled water

Reagent preparation

① Equilibrate reagents to room temperature before use.

② The preparation of chromogenic agent:

Dissolve one vial of chromogenic agent A and one vial of chromogenic agent B with 15 mL of chromogenic agent C, mix well to dissolve. Store at 2-8°C for 1 month protected from light.

③ The preparation of standard curve:

Always prepare a fresh set of standards. Discard working standard dilutions after use.

Dilute 100 mg/L iron standard stock solution with double distilled water to a serial concentration. The recommended dilution gradient is as follows: 0, 5, 10, 20, 25, 30, 40, 50 mg/L. Reference is as follows:

Item	①	②	③	④	⑤	⑥	⑦	⑧
Concentration (mg/L)	0	5	10	20	25	30	40	50
100 mg/L standard (μL)	0	10	20	40	50	60	80	100
Double distilled water (μL)	200	190	180	160	150	140	120	100

Sample preparation

① Sample preparation

Serum and plasma: detect directly. If not detected on the same day, the serum or plasma can be stored at -80°C for a month.

② Dilution of sample

The recommended dilution factor for different samples is as follows (for reference only):

Sample type	Dilution factor
Human serum	1
Rat serum	1
Porcine serum	1
Rabbit serum	1
Chicken serum	1
Machin serum	1

Note: The diluent is double distilled water. For the dilution of other sample types, please do pretest to confirm the dilution factor

The key points of the assay

- ① After 100°C water bath, the supernatant obtained by centrifugation must be clarified, otherwise the experimental results will be affected.
- ② The experimental container must be clean to avoid the contamination of iron.

Operating steps

The measurement of standard curve:

- ① Dilute 100 mg/L iron standard stock solution with double distilled water to a serial concentration. The recommended dilution gradient is as follows: 0, 5, 10, 20, 25, 30, 40, 50 mg/L.
- ② Take 30 μ L of standard solution with different concentration to the wells.
- ③ Add 150 μ L of chromogenic agent to the wells.
- ④ Mix fully for 5 s with microplate reader, stand at room temperature for 5 min and measure the OD value at 520 nm.

The measurement of sample:

- ① The pretreatment of sample
Take 50 μ L of serum, add 50 μ L of 10 mg/L iron standard application solution, mix fully with a vortex mixer and stand at room temperature for 5 min. Then add a vial of iron absorbent, mix fully with a vortex mixer for 3 s and stand at room temperature for 5 min. Centrifuge at 3000 \times g for 10 min and take the supernatant for detection
- ② Sample tube: Add 50 μ L of pretreated sample into the 1.5 mL EP tube.
Control tube: Add 50 μ L of double distilled water into the 1.5 mL EP tube.
- ③ Add 250 μ L of chromogenic agent into each tube. Oscillate fully with a vortex mixer for 3 s and incubate in 100 $^{\circ}$ C water bath for 5 min.
- ④ Cool the tubes with running water, then centrifuge at 10000 \times g for 10 min (If the supernatant is turbid, collect the turbid supernatant into another new EP tube and centrifuge again).
- ⑤ Take 180 μ L of the supernatant to the corresponding wells of microplate and measure the OD value at 520 nm of each well.

Calculation

The standard curve:

1. Average the duplicate reading for each standard.
2. Subtract the mean OD value of the blank (Standard # ①) from all standard readings. This is the absolved OD value.
3. Plot the standard curve by using absolved OD value of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve ($y = ax + b$) with graph software (or EXCEL).

The sample:

1. Serum (plasma) sample:

$$\text{TIBC (mg/L)} = (\Delta A_{520} - b) \div a \times f$$

or

$$\text{TIBC (}\mu\text{mol/L)} = (\Delta A_{520} - b) \div a \times f \times c_1$$

$$\text{UIBC}(\mu\text{mol/L)} = c_3 - c_2$$

$$i = c_2 \div c_3 \times 100 \%$$

[Note]

f: Dilution factor of sample before test.

ΔA_{520} : $OD_{\text{Sample}} - OD_{\text{Blank}}$.

c_1 : 17.91 $\mu\text{mol/L}$ (1 mg/L Iron = 17.91 $\mu\text{mol/L}$).

c_2 : The concentration of serum iron.

c_3 : Total iron binding capacity (TIBC) ($\mu\text{mol/L}$).

i: Iron saturation (%).

Appendix I Performance Characteristics

1. Parameter:

Intra-assay Precision

Three human serum samples were assayed in replicates of 20 to determine precision within an assay (CV = Coefficient of Variation).

Parameters	Sample 1	Sample 2	Sample 3
Mean (mg/L)	3.80	24.60	43.80
%CV	1.7	1.6	1.2

Inter-assay Precision

Three human serum samples were assayed 20 times in duplicate by three operators to determine precision between assays.

Parameters	Sample 1	Sample 2	Sample 3
Mean (mg/L)	3.80	24.60	43.80
%CV	2.5	2.1	2.3

Recovery

Take three samples of high concentration, middle concentration and low concentration to test the samples of each concentration for 6 times parallelly to get the average recovery rate of 100%.

	Standard 1	Standard 2	Standard 3
Expected Conc. (mg/L)	8.5	22.6	32
Observed Conc. (mg/L)	8.6	22.4	32.0
recovery rate(%)	101	99	100

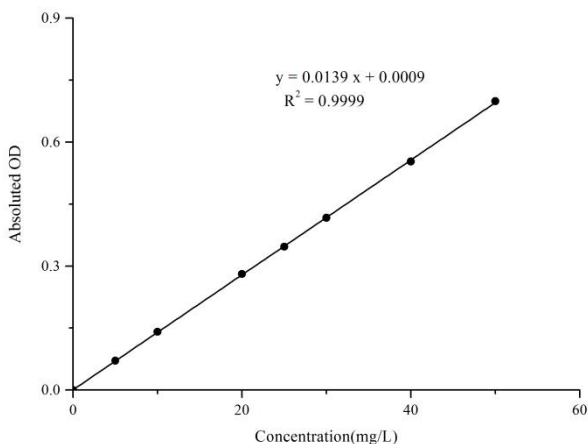
Sensitivity

The analytical sensitivity of the assay is 0.14 mg/L. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 20 times, and calculating the corresponding concentration.

2. Standard curve:

As the OD value of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique or temperature effects), so the standard curve and data are provided as below for reference only:

Concentration (mg/L)	0	5	10	20	25	30	40	50
Average OD	0.037	0.108	0.178	0.318	0.384	0.454	0.590	0.736
Absoluted OD	0	0.071	0.141	0.281	0.347	0.417	0.553	0.699



Appendix Π Example Analysis

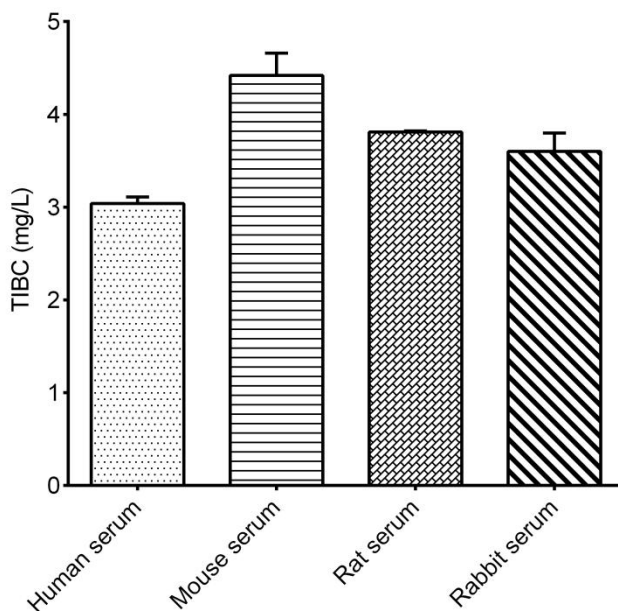
Example analysis :

Take 50 μL of human serum and carry the assay according to the operation steps. The results are as follows:

Standard curve: $y = 0.014x - 0.0009$, the average OD value of the sample is 0.080, the average OD value of the control is 0.038, and the calculation result is:

$$\text{TIBC (mg/L)} = (0.080 - 0.038 + 0.0009) \div 0.014 = 3.06 \text{ mg/L}$$

Detect human serum, mouse serum, rat serum and rabbit serum according to the protocol, the result is as follows:



Statement

1. This assay kit is for Research Use Only. We will not response for any arising problems or legal responsibilities causing by using the kit for clinical diagnosis or other purpose.
2. Please read the instructions carefully and adjust the instruments before the experiments. Please follow the instructions strictly during the experiments.
3. Protection methods must be taken by wearing lab coat and latex gloves.
4. If the concentration of substance is not within the detection range exactly, an extra dilution or concentration should be taken for the sample.
5. It is recommended to take a pre-test if your sample is not listed in the instruction book.
6. The experimental results are closely related to the situation of reagents, operations, environment and so on. Elabscience will guarantee the quality of the kits only, and NOT be responsible for the sample consumption caused by using the assay kits. It is better to calculate the possible usage of sample and reserve sufficient samples before use.

