（FOR RESEARCH USE ONLY. DO NOT USE IT IN CLINICAL DIAGNOSIS !）

Catalog No：E-BC-K784-M

Specification：96T(40 samples)

Measuring instrument：Microplate reader(440-460 nm)

Detection range：0.11-8.33 U/L

Elabscience® Fatty Acid Oxidation (FAO) Colorimetric Assay Kit

This manual must be read attentively and completely before using this product.
If you have any problem, please contact our Technical Service Center for help：

Toll-free: 1-888-852-8623
Tell: 1-832-243-6086
Fax: 1-832-243-6017
Email: techsupport@elabscience.com
Website: www.elabscience.com

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

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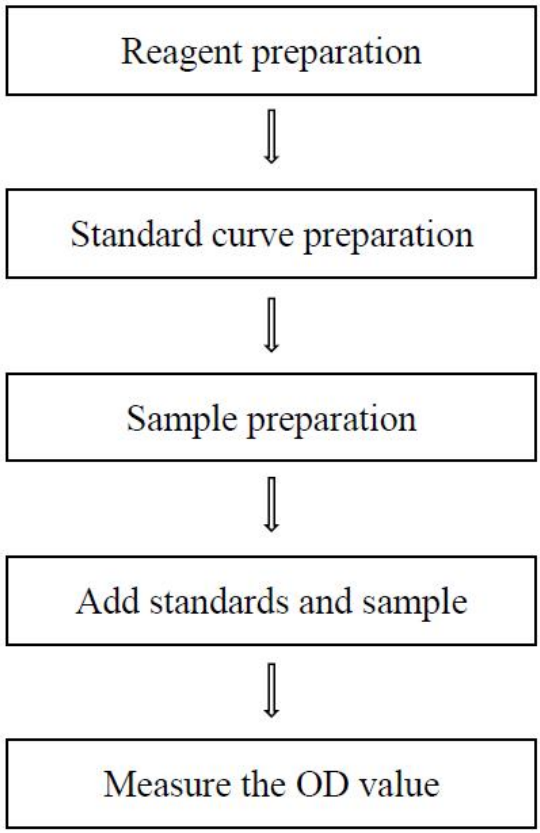
Assay summary



Intended use

This kit can be used to measure fatty acid oxidation (FAO) ability in animal tissue and cell samples.

Detection principle

Fatty acid oxidation (FAO) is the main pathway of fatty acid decomposition in the body, which can supply a large amount of energy required by the organism. FAO is also the transformation process of fatty acids. The length of fatty acid chain required by the organism is different. Through oxidation, long-chain fatty acids can be transformed into fatty acids with suitable growth degree for the organism metabolism.

The detection principle of this kit: FAO process consumes the substrate and NAD^+, and generates NADH. Under the action of electron coupling agent and chromogenesis agent, the orange red substance is generated, which can be detected at 450 nm.

Kit components & storage

Item	Component	Size (96 T)	Storage
Reagent 1	Buffer Solution	40 mL × 1 vial	-20°C, 12 months, shading light
Reagent 2	Co-factor	Powder ×2 vials	-20°C, 12 months, shading light
Reagent 3	Substrate	0.22 mL ×1 vial	-20°C, 12 months, shading light
Reagent 4	Chromogenic Agent	1.2 mL ×2 vials	-20°C, 12 months, shading light
Reagent 5	Standard	Powder ×2 vials	-20°C, 12 months, shading light
	Microplate	96 wells	No requirement
	Plate Sealer	2 pieces	

Note: The reagents must be stored strictly according to the preservation conditions

in the above table. The reagents in different kits cannot be mixed with each other. For a small volume of reagents, please centrifuge before use, so as not to obtain sufficient amount of reagents.

Materials prepared by users

Instruments:

Microplate reader (440-460 nm, optimum wavelength: 450 nm), Incubator (37℃)

Reagents:

Normal saline (0.9% NaCl)

Reagent preparation

① Equilibrate all the reagents to 25°C before use.

② Preparation of Co-factor working solution：
Dissolve one vial of Co-factor with 1 mL of buffer solution, mix well to dissolve. Keep it on ice during use protected from light, store at -20°C protected from light and used up within 3 days.

③ Preparation of substrate working solution：
Before testing, please prepare sufficient substrate working solution according to the test wells. For example, prepare 50 μL of substrate working solution (mix well 40 μL of buffer solution and 10 μL of substrate). The substrate working solution should be prepared on spot and keep it on ice during use protected from light. Store at -20°C protected from light and used up within 3 days.

④ Preparation of reaction working solution：
For each well, prepare 154 μL of reaction working solution (mix well 144 μL of buffer solution and 10 μL of Co-factor working solution). The reaction working solution should be prepared on spot. Keep it on ice during use protected from light and used up on the same day.

⑤ Preparation of 0.5 mmol/L standard solution：

Dissolve one vial of standard with 5 mL of double distilled water, mix well to dissolve. Keep it on ice during use protected from light. Store at -20°C protected from light and used up within 3 days.

⑥ The preparation of standard curve：

Always prepare a fresh set of standards. Discard working standard dilutions after use.

Dilute 0.5 mmol/L standard solution with double distilled water to a serial concentration. The recommended dilution gradient is as follows: 0, 0.05, 0.1, 0.125, 0.15, 0.175, 0.2, 0.25 mmol/L. Reference is as follows:

Item	①	②	③	④	⑤	⑥	⑦	⑧
Concentration (mmol/L)	**0**	**0.1**	**0.15**	**0.2**	**0.3**	**0.4**	**0.45**	**0.5**
0.5 mmol/L standard (μL)	0	40	60	80	120	160	180	200
Double distilled water (μL)	200	160	140	120	80	40	20	0

Sample preparation

① Sample preparation

Tissue sample:

① Harvest the amount of tissue needed for each assay (initial recommendation 30 mg).

② Wash tissue in normal saline.

③ Homogenize 30 mg tissue in 270 μL normal saline with a dounce homogenizer at 4℃.

④ Centrifuge at 10000×g for 15 min to remove insoluble material. Collect supernatant and keep it on ice for detection. Detect the prepared tissue supernatant within 4 h.

⑤ Meanwhile, determine the protein concentration of supernatant (E-BC-K318-M).

Cell sample:

① Harvest the number of cells needed for each assay (initial recommendation 1×10^6 cells).

② Wash cells with normal saline.

③ Homogenize 1×10^6 cells in 200 μL normal saline with a ultrasonic cell disruptor at 4℃.

④ Centrifuge at 10000 × g for 15 min at 4℃ to remove insoluble material. Collect supernatant and keep it on ice for detection. Detect the prepared tissue supernatant within 4 h.

⑤ Meanwhile, determine the protein concentration of supernatant (E-BC-K318-M).

② Dilution of sample

The recommended dilution factor for different samples is as follows (for reference only):

Sample type	Dilution factor
10% Mouse liver tissue homogenate	2-5
10% Mouse brain tissue homogenate	1
10% Mouse lung tissue homogenate	1
10% Mouse spleen tissue homogenate	1
10% Mouse kidney tissue homogenate	2-5
10% Mouse heart tissue homogenate	2-3
10% Mouse muscle tissue homogenate	1-3
1×10^6 Molt-4 cells	1
1×10^6 HL-60 cells	1
1×10^6 Jurkat cells	1

Note: The diluent is normal saline (0.9% NaCl). For the dilution of other sample types, please do pretest to confirm the dilution factor.

The key points of the assay

If the OD value of the sample is greater than 1.5, dilute the sample.

Operating steps

① Standard well: Add 50 μL of standard solution with different concentrations to the corresponding wells.

Sample well: Add 50 μL of sample to the corresponding wells.

Control well: Add 50 μL of sample to the corresponding wells.

② Add 165 μL of buffer solution to standard wells, add 20 μL of substrate working solution to sample wells and add 20 μL of buffer solution to control wells.

③ Add 145 μL of reaction working solution to sample and control wells.

④ Add 20 μL of chromogenic agent to each well.

⑤ Mix fully with microplate reader for 5 s and incubate at 37℃ for 30 min. Measure the OD value of each well at 450 nm with microplate reader.

Calculation

The standard curve:

1. Average the duplicate reading for each standard.

2. Subtract the mean OD value of the blank (Standard #①) from all standard readings. This is the absoluted OD value.

3. Plot the standard curve by using absoluted OD value of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve (**y = ax + b**) with graph software (or EXCEL).

The sample:

Tissue and cell samples:

Definition: The amount of enzyme in 1 g tissue or cell protein per 1 minute that hydrolyze the substrate to produce 1 μmol NADH at 37°C is defined as 1 unit.

$$\text{FAO ability(U/gprot)} = (\Delta A_{450} - b) \div a \div T \times 1000 \div C_{pr} \times f$$

[Note]

ΔA_{450}: $OD_{Sample} - OD_{Control}$.

T: Reaction time, 30 min.

1000*: 1 mmol/L = 1000 μmol/L.

C_{pr}: The concentration of protein in sample, gprot/L.

f: Dilution factor of sample before test.

Appendix I Performance Characteristics

1. Parameter:

Intra-assay Precision

Three mouse liver samples were assayed in replicates of 20 to determine precision within an assay (CV = Coefficient of Variation).

Parameters	Sample 1	Sample 2	Sample 3
Mean (U/L)	1.00	3.00	6.00
%CV	4.1	4.8	5.0

Inter-assay Precision

Three mouse liver samples were assayed 20 times in duplicate by three operators to determine precision between assays.

Parameters	Sample 1	Sample 2	Sample 3
Mean (U/L)	1.00	3.00	6.00
%CV	8.2	9.1	9.4

Recovery

Take three samples of high concentration, middle concentration and low concentration to test the samples of each concentration for 6 times parallelly to get the average recovery rate of 104%.

	Sample 1	Sample 2	Sample 3
Expected Conc. (U/L)	1.00	3.00	6.00
Observed Conc. (U/L)	1.0	3.0	6.6
Recovery rate (%)	102	100	110

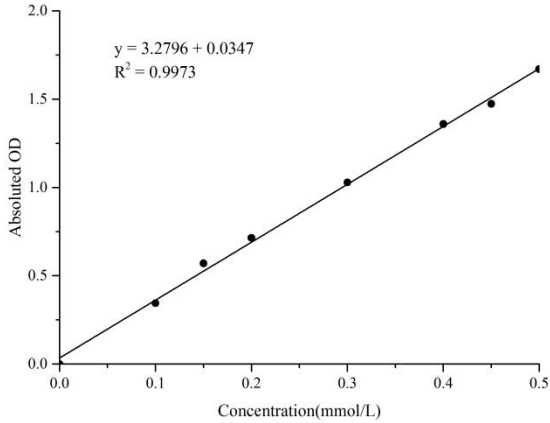
Sensitivity

The analytical sensitivity of the assay is 0.11 U/L. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 20 times, and calculating the corresponding concentration.

2. Standard curve:

As the OD value of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique or temperature effects), so the standard curve and data are provided as below for reference only:

Concentration (mmol/L)	0	0.1	0.15	0.2	0.3	0.4	0.45	0.5
OD	0.05	0.425	0.615	0.763	1.003	1.41	1.534	1.716
	0.05	0.365	0.626	0.766	1.154	1.41	1.513	1.726
Average OD	0.050	0.395	0.621	0.765	1.079	1.410	1.524	1.721
Absoluted OD	0.000	0.345	0.571	0.715	1.029	1.360	1.474	1.671



Appendix II Example Analysis

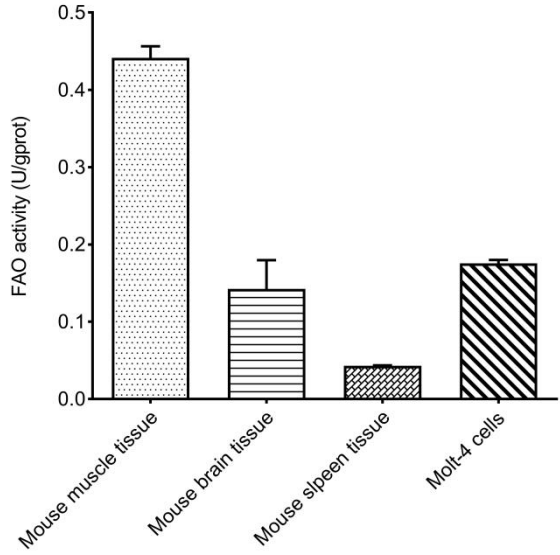
Example analysis：

Take 50 μL of 10% mouse muscle tissue homogenate, and carry the assay according to the operation steps. The results are as follows:

standard curve: y = 3.2796 x+0.0347, the average OD value of the sample well is 1.860, the average OD value of the control well is 1.600, ΔA_{450} = 1.860 - 1.600 = 0.260, the concentration of protein is 5.201 gprot/L, and the calculation result is:

FAO ability(U/grpot) = (0.260 - 0.0347) ÷ 3.2796 ÷ 30 × 1000 ÷ 5.201 = 0.44 U/gprot

Detect 10% mouse muscle tissue homogenate (the concentration of protein is 5.201 gprot/L), 10% mouse brain tissue homogenate (the concentration of protein is 3.931 gprot/L), 10% mouse spleen tissue homogenate (the concentration of protein is 8.297 gprot/L), 1×10^6 Molt-4 cells homogenate (the concentration of protein is 0.165 gprot/L), according to the protocol, the result is as follows：



Statement

1. This assay kit is for Research Use Only. We will not response for any arising problems or legal responsibilities causing by using the kit for clinical diagnosis or other purpose.
2. Please read the instructions carefully and adjust the instruments before the experiments. Please follow the instructions strictly during the experiments.
3. Protection methods must be taken by wearing lab coat and latex gloves.
4. If the concentration of substance is not within the detection range exactly, an extra dilution or concentration should be taken for the sample.
5. It is recommended to take a pre-test if your sample is not listed in the instruction book.
6. The experimental results are closely related to the situation of reagents, operations, environment and so on. Elabscience will guarantee the quality of the kits only, and NOT be responsible for the sample consumption caused by using the assay kits. It is better to calculate the possible usage of sample and reserve sufficient samples before use.