

#### **BCA Protein Colorimetric Assay Kit**

Catalog No:	E-BC-K318-M Method:		Colorimetric method					
Instrument:	Microplate reader(540-590 nm)	Specification:	96T/ 500Assays					
	<ol> <li>Note: 1 The samples should not contain EGTA, EDTA and other chelating agents, and should not contain DTT, mercaptoethanol and other reducing substances.</li> <li>2 Protein concentration: When the sample background value is high, the bradford protein colorimetric assay kit is recommended (E-BC-K168-S); When the protein concentration is high, the biuret protein colorimetric assay kit is recommended (E-BC-K165-S).</li> </ol>							
General information								
Intended use	This kit can be used to me	easure total protein	(TP) content in serum, plasma, culture cells,					

tissue and cells samples.

**Detection range** Detection range: 0.0165-1 mg/mL

and sensitivity Sensitivity: 0.0165 mg/mL

**Detection principle** Cu<sup>2+</sup> can be reduced to Cu<sup>+</sup> by protein in alkaline condition. Cu<sup>+</sup> can combine with BCA reagent and form purple complex, which has a maximum absorption peak at 562 nm. The absorbance value is proportional to the protein concentration. Therefore, the protein concentration can be calculated according to the OD value.

Item	Component	Size 1 (96 T)	Size 2 (500 Assays)	Storage	
Reagent 1	BCA Reagent	$25 \text{ mL} \times 1 \text{ vial}$	$50 \text{ mL} \times 2 \text{ vials}$	RT, 12 months	
Reagent 2	Copper Salt Solution	$0.5 \text{ mL} \times 1 \text{ vial}$	$3 \text{ mL} \times 1 \text{ vial}$	RT, 12 months	
Reagent 3	Standard	$1 \text{ mg} \times 1 \text{ vial}$	$1 \text{ mg} \times 5 \text{ vials}$	RT, 12 months	
Reagent 4	Standard Diluent	15 mL × 1 vial	$30 \text{ mL} \times 1 \text{ vial}$	RT, 12 months	
	Microplate	96 wells	/	No requirement	
	Plate Sealer	2 pieces			

#### Kit components & storage

Note: The reagents must be stored strictly according to the preservation conditions in the above table.

The reagents in different kits cannot be mixed with each other.

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# Materials prepared by users

#### Instruments:

Microplate reader (540-590 nm), Vortex mixer, Micropipettor, Incubator

#### **Reagents:**

Double distilled water, Normal saline (0.9% NaCl), PBS (0.01 M, pH 7.4)

# **Reagent preparation**

① The preparation of 1 mg/mL standard solution:

Dissolve a vial of standard with 1 mL standard diluent and mix fully. Aliquoted storage at -20°C for 3 months.

2 The preparation of BCA working solution:

For each well, prepare 200  $\mu$ L of BCA working solution (mix well 4  $\mu$ L of copper salt solution and 196  $\mu$ L of BCA reagent). Store at 2-8°C for 24 h.

(3) The preparation of standard curve:

Always prepare a fresh set of standards. Discard working standard dilutions after use.

Dilute 1 mg/mL standard solution with standard diluent to a serial concentration. The recommended dilution gradient is as follows: 0, 0.1, 0.2, 0.3, 0.4, 0.6, 0.8, 1.0 mg/mL. Reference is as follows.

Item		2	3	4	5	6	7	8
Concentration (mg/mL)		0.1	0.2	0.3	0.4	0.6	0.8	1.0
1 mg/mL standard solution (μL)		20	40	60	80	120	160	200
Standard Diluent (µL)		180	160	140	120	80	40	0

# **Operation steps**

1 Standard well: add 20  $\mu L$  of standard solution with different concentration.

Sample well: add 20 µL of tested samples.

- 2 Add 200  $\mu L$  of BCA working solution into each tube.
- ③ Oscillate for 20 s to mix fully and incubate at 37°C for 30 min.
- ④ Measure the OD values of each well at 562 nm with microplate reader.
- Note: When the reagent is added to the wells, it should be added to the bottom of the enzyme well; Add the sample slowly to avoid bubbles (Bubble influence test results).

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# Calculation

### The standard curve:

- 1. Average the duplicate reading for each standard.
- 2. Subtract the mean OD value of the blank (Standard #1) from all standard readings. This is the absoluted OD value.
- 3. Plot the standard curve by using absoluted OD value of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve (y = ax + b) with graph software (or EXCEL).

## The sample:

Protein content (mg/mL) =  $(\Delta A_{562} - b) \div a \times f$ 

### Note:

 $\Delta A_{562}$ : Absolute OD (OD<sub>Sample</sub> - OD<sub>Blank</sub>).

f: Dilution factor of sample before test.