

MG (Malachite Green) ELISA Kit

Catalog No: E-FS-E180

96T/96T*3

Version Number:	V1.1
Replace version:	V1.0
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This manual must be read attentively and completely before using this product.

If you have any problems, please contact our Technical Service Center for help.

Toll-free: 1-888-852-8623 Tel: 1-832-243-6086 Fax: 1-832-243-6017

Email: techsupport@elabscience.com

Website: www.elabscience.com

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

Test principle

This kit uses Competitive-ELISA as the method for the quantitative detection. It can detect Malachite Green (MG) in samples, such as fish, shrimp, bullfrog, etc. This kit is composed of ELISA Microtiter plate, HRP conjugate, antibody working solution, standard and other supplementary reagents. The microtiter plate in this kit has been pre-coated with coupled antigen. During the reaction, MG in the samples or standard competes with coupled antigen on the solid phase supporter for sites of anti-MG antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each microtiter plate well, and substrate reagent is added for color development. There is a negative correlation between the OD value of samples and the concentration of MG. The concentration of MG in the samples can be calculated by comparing the OD of the samples to the standard curve.

Technical indicators

Reaction mode (Incubation time and temperature): 25±2°C; 30 min, 15-20 min.

Detection limit: 0.5 ppb.

Cross-reactivity: Explicit Malachite Green ---100%; Hidden Malachite Green (oxidized)---100%;
Explicit Crystal Violet--- 77%; Hidden Crystal Violet (oxidized) --- 77%.

Sample recovery rate: 95%±25%.

Kit components

Item	Specifications
ELISA Microtiter plate	96 wells
20×Concentrated Standard Liquid	1 mL each (ppb=ng/mL=ng/g) (0 ppb, 1 ppb, 3 ppb, 9 ppb, 27 ppb, 81 ppb)
Standard Diluent	50 mL
HRP Conjugate	7 mL
Antibody Working Solution	7 mL
4×Concentrated Sample Diluent A	15 mL
Purification Reagent	100 g
10×Concentrated Oxidizing Solution	1 mL
Sample Diluent B	100 mL*2
20×Concentrated Wash Buffer	25 mL
Substrate Reagent A	7 mL
Substrate Reagent B	7 mL
Stop Solution	7 mL
Plate Sealer	1 piece
Sealed Bag	1 piece
Manual	1 copy

Note: All reagent bottle MGs must be tightened to prevent evaporation and microbial pollution

Other materials required but not supplied

Instrument: Microplate reader, Homogenizer, Nitrogen evaporators, Water bath, Vortex mixer, Centrifuge, Graduated pipette, Balance (sensitivity 0.01 g).

Micropipette: Single channel (20-200 μ L, 100-1000 μ L), Multichannel (30-300 μ L).

Reagents: Acetonitrile.

Notes

1. The overall OD value will be lower when reagents have not been brought to room temperature before use or room temperature is below $25\pm 2^{\circ}\text{C}$.
2. If the wells turn dry during the washing procedure, it will lead to bad linear standard curve and poor repeatability. Operate the next step immediately after wash.
3. Mix thoroughly and wash the plate completely. The consistency of wash procedure can strongly affect the reproducibility of this ELISA kit.
4. FOR RESEARCH USE ONLY. ELISA Microtiter plate should be covered by plate sealer. Avoid the kit to strong light.
5. **Each reagent is optimized for use in the E-FS-E180. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other E-FS-E180 with different lot numbers.**
6. Substrate Reagent should be abandoned if it turns color. When OD value of standard (concentration: 0) < 0.8 unit ($A_{450\text{nm}} < 0.8$), it indicates the reagent may be deteriorated.
7. Stop solution is caustic, avoid contact with skin and eyes.
8. As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.
9. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
10. **For mentioned sample fast and efficient extraction methods are included in the kit description. Please consult technical support for the applicability if other sample need to be tested.**
11. The time of each step of adding the sample solution, enzyme conjugate working solution, AB mixture and stop solution in plate well should not exceed 3 min.
12. The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS, etc. can be used for quantitative confirmation.

Storage and expiry date

Store the kit at $2-8^{\circ}\text{C}$. Do not freeze any test kit components.

Return any unused microwells to their original foil bag and reseal them together with the desiccant provided and further store at $2-8^{\circ}\text{C}$. After opening, the kit is stable for up to 1 month.

Expiry date: expiration date is on the packing box

Experimental preparation

Restore all reagents and samples to room temperature ($25\pm 2^{\circ}\text{C}$) before use.

Open the microplate reader in advance, preheat the instrument, and set the testing parameters.

1. Sample pretreatment Notice:

Experimental apparatus should be clean, and the pipette should be disposable to avoid cross-contamination during the experiment.

2. Solution preparation

Please prepare solution according to the number of samples. Don't use up all components in the kit at once!

Solution 1: **Sample Diluent A**

Dilute the **4×Concentrated Sample Diluent A** with deionized water. (4×Concentrated Sample Diluent A (V): Deionized water (V) = 1:3).

Solution 2: **Oxidizing Solution**

Dilute the **10×Concentrated Oxidizing Solution** with acetonitrile. (10×Concentrated Oxidizing Solution (V): Acetonitrile (V) = 1:9).

Prepare the fresh solution before use and keep the unused solution for no more than 2 hours.

Solution 3: **Wash Buffer**

Dilute **20×Concentrated Wash Buffer** with deionized water (20×Concentrated Wash Buffer (V): Deionized water (V) = 1:19).

Solution 4: **Standard Liquid**

Dilute **20×Concentrated Standard Liquid** with **Standard Diluent**. (20×Concentrated Standard Liquid (V): Standard Diluent (V) = 1:19).

Prepare the fresh solution before use and keep the unused solution for no more than 24 hours.

3. Sample pretreatment

3.1 Pretreatment of fish, shrimp, bullfrog sample:

- (1) Remove fat from sample. Homogenize the representative sample with a homogenizer and mix fully.
- (2) Weigh 1 ± 0.05 g of homogenate sample into a 50 mL centrifuge tube, add 0.5 mL of **Sample Diluent A** (Solution 1) and 4 mL of acetonitrile, vortex for 3 min. Centrifuge at 4000 g for 5 min at room temperature.
- (3) Take 1.0 g of **Purification Reagent** to a new 5 mL centrifuge tube, then add 2 mL of the supernatant, vortex for 1 min, standing at room temperature for 5 min. Centrifuge at 4000 g for 5 min at room temperature.
- (4) Take 30 μ L of **Oxidizing Solution** (Solution 2) to a new 5 mL centrifuge tube, then add 1 mL of the supernatant, vortex for 1 min, dry at 50-60°C with nitrogen evaporators. (Please do it in a ventilated environment.)
- (5) Dissolve the residue with 2 mL of **Sample Diluent B** and vortex for 1 min. Store away from light for future use.
- (6) Take 50 μ L for analysis.

Note: Sample dilution factor: 10 detection limit: 0.5 ppb

Assay procedure

Restore all reagents and samples to room temperature before use. All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming. The unused ELISA Microtiter plate should be sealed as soon as possible and stored at 2-8°C.

1. **Number:** number the sample and standard in order (multiple well), and keep a record of standard wells and sample wells. **Standard and Samples need test in duplicate**
2. **Add Sample:** add 50 μ L of **Standard Liquid** (Solution 4) or **Sample** per well, add 50 μ L **HRP Conjugate**, then add 50 μ L of **Antibody Working Solution**, cover the plate with plate sealer. Oscillate for 10 s gently to mix thoroughly. Incubate at $25\pm 2^\circ\text{C}$ for 30 min in shading light.
3. **Wash:** uncover the sealer carefully, remove the liquid in each well. Immediately add 260 μ L of **Wash Buffer** (Solution 3) to each well. Repeat wash procedure for 4 times, 15-30 s intervals/time. Invert the plate and pat it against thick clean absorbent paper (If bubbles exist in the wells, clean tips can be used to prick them).
4. **Color Development:** add 100 μ L of **Substrate mixed solution** to each well (**Substrate Reagent A** and **Substrate Reagent B** are fully mixed at ratio 1:1 by volume, the mixture should be used within 5 min, avoid using metal containers or stirring the reagents), Gently oscillate for 10 s to mix thoroughly. Incubate at $25\pm 2^\circ\text{C}$ for 15-20 min in shading light.
5. **Stop Reaction:** add 50 μ L of **Stop Solution** to each well. Gently oscillate for 10 s to mix thoroughly.
6. **OD Measurement:** determine the optical density (OD value) of each well at 450 nm (reference wavelength 630 nm) with a microplate reader. This step should be finished in 5 min after stop reaction.

Result analysis

1. Absorbance (%)= $A/A_0 \times 100\%$

A: Average absorbance of standard or sample

A_0 : Average absorbance of 0 ppb Standard

2. Drawing and calculation of standard curve

Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add average absorbance value of sample to standard curve to get corresponding concentration. **If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.**

For this kit, it is more convenient to use professional analysis form for accurate and fast analysis on a large number of samples.

Malachite Green (E-FS-E180) Standard Curve

