(FOR RESEARCH USE ONLY, DO NOT USE IT IN CLINICAL DIAGNOSIS!)

Catalog No: E-BC-K064-M

Specification: 96T(40 samples)

Measuring instrument: Microplate reader (390-415 nm)

Detection range: 0.76-49.51 U/L

Elabscience®β-N-Acetylglucosaminidase (NAG) Activity Assay Kit

This manual must be read attentively and completely before using this product.

If you have any problem, please contact our Technical Service Center for help:

Toll-free: 1-888-852-8623

Tell: 1-832-243-6086

Fax: 1-832-243-6017

Email: techsupport@elabscience.com

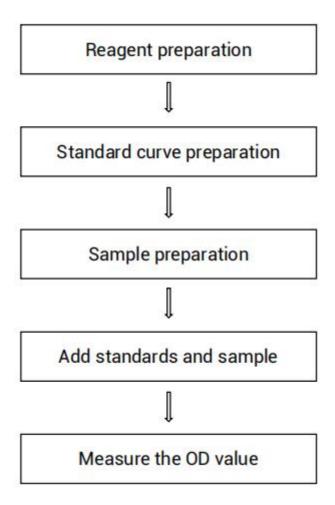
Website: www.elabscience.com

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

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Assay summary



Intended use

This kit can measure β -N-Acetylglucosaminidase (NAG) activity in serum, plasma, animal tissue and cell samples.

Detection principle

β-N-Acetylglucosaminidase (NAG) catalyzed the substrate to produce p-nitrophenol, which has the maximum absorption peak at about 400 nm. Therefore, the activity of NAG can be calculated by measuring the change of absorbance value at 400 nm.

Kit components & storage

Item	Component	Size (96 T)	Storage
Reagent 1	Buffer Solution	20 mL × 1 vial	-20℃, 12 months
Reagent 2	Substrate Solution	5 mL × 1 vial	-20℃, 12 months, shading light
Reagent 3	Chromogenic Agent	14 mL × 1 vial	-20℃, 12 months
Reagent 4	Standard	Powder × 1 vial	-20℃, 12 months, shading light
	Microplate	96 wells	No requirement
	Plate Sealer	2 pieces	
	Sample Layout Sheet	1 piece	

Note: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other. For a small volume of reagents, please centrifuge before use, so as not to obtain sufficient amount of reagents.

Materials prepared by users

Instruments:

Microplate reader (390-415 nm, optimum wavelength: 400 nm), Incubator(37 $^{\circ}$ C)

Reagents:

Double distilled water, Normal saline (0.9% NaCl)

Reagent preparation

- ① Equilibrate all reagents to room temperature before use
- 2 The preparation of measure working solution: Before testing, please prepare sufficient mersuring working solution according to the test wells. For example, prepare 180 μ L measure working solution(add 140 μ L of buffer solution and 40 μ L of substrate solution, mix well). The measure working solution should be prepared on spot. The prepared solution should be used up within 1 day.
- ③ The preparation of 20 mmol/L standard solution:

 Dissolve one vial of standard with 1 mL of double distilled water and heat at 40℃ for 3 min protected from light until dissolved. Aliquoted storage at -20°C for 3 months.
- ④ The preparation of 0.5 mmol/L standard solution: Dilute 20 μL of 20 mmol/L standard solution with 780 μL of double distilled water, mix well. The enzyme working solution should be prepared on spot.
- (5) The preparation of standard curve:

 Always prepare a fresh set of standards. Discard working standard dilutions after use.
 - Dilute 0.5 mmol/L standard with double distilled water to a serial concentration. The recommended dilution gradient is as follows: 0, 0.1,

0.15, 0.2, 0.3, 0.35, 0.4, 0.5 mmol/L. Reference is as follows:

Item	1	2	3	4	(5)	6	7	8
Concentration (mmol/L)	0	0.1	0.15	0.2	0.3	0.35	0.4	0.5
0.5 mmol/L standard (μL)	0	40	60	80	120	140	160	200
Double distilled water (µL)	200	160	140	120	80	60	40	0

Sample preparation

① Sample preparation

Serum and plasma: detect directly. If not detected on the same day, the serum or plasma can be stored at -80° C for a month.

Tissue sample:

- ① Harvest the amount of tissue needed for each assay (initial recommendation 20 mg).
- 2 Wash tissue in cold PBS (0.01 M, pH 7.4).
- ③ Homogenize 20 mg tissue in 180 μL normal saline(0.9% NaCl) with a dounce homogenizer at 4° C.
- 4 Centrifuge at 10000×g for 10 minutes at 4 $^{\circ}$ C to remove insoluble material. Collect supernatant and keep it on ice for detection.
- (E-BC-K318-M).

Cell (adherent or suspension) samples:

- ① Harvest the number of cells needed for each assay (initial recommendation 1×10⁶ cells).
- ② Wash cells with PBS (0.01 M, pH 7.4).
- ③ Homogenize 1×10⁶ cells in 200 μ L normal saline(0.9% NaCl) with a ultrasonic cell disruptor at 4°C.
- ④ Centrifuge at 10000×g for 10 minutes at 4 °C to remove insoluble

material. Collect supernatant and keep it on ice for detection.

(5) Meanwhile, determine the protein concentration of supernatant(E-BC-K318-M).

2 Dilution of sample

The recommended dilution factor for different samples is as follows (for reference only):

Sample type	Dilution factor
10% Mouse liver tissue homogenate	3-5
10% Mouse kidney tissue homogenate	3-5
10% Mouse heart tissue homogenate	3-5
10% Mouse lung tissue homogenate	3-5
Rat plasma	1
Human serum	1
Bovine serum	2-3
6×10^6 CHO cells	1
0.6×10^6 293T/17 cells	1

Note: The diluent is normal saline (0.9% NaCl). For the dilution of other sample types, please do pretest to confirm the dilution factor.

Operating steps

① Standard well: Add 20 μL of different concentrations solution to standard well.

Sample well: Add 20 μL of sample to sample well.

Control well: Add 20 µL of sample to control well.

- ② Add 160 μ L of measure working solution to standard well and sample well. Add 160 μ L of buffer solution to control well.
- ③ Mix fully with microplate reader for 5 s, incubated at 37°C for 10 min.
- 4 Add 100 µL of chromogenic agent to each well.
- S Mix fully with microplate reader for 5 s, Measure the OD value of each well at 400 nm with microplate reader.

Calculation

The standard curve:

- 1. Average the duplicate reading for each standard.
- 2. Subtract the mean OD value of the blank (Standard # ①) from all standard readings. This is the absoluted OD value.
- 3. Plot the standard curve by using absoluted OD value of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve (y = ax + b) with graph software (or EXCEL).

The sample:

1. Serum (plasma) sample:

Definition: The amount of NAG in 1 L liquid sample per 1 minute that hydrolyze the substrate to produce 1 μ mol product at 37 $^{\circ}$ C is defined as 1 unit

NAG activity
(U/L) =
$$(\Delta A_{400} - b) \div a \div T \times f \times 1000$$

2. Tissue sample:

Definition: The amount of NAG in 1 g tissue or cell per 1 minute that hydrolyze the substrate to produce 1 μ mol product at 37 $^{\circ}$ C is defined as 1 unit.

NAG activity (U/gprot) =
$$(\Delta A_{400} - b) \div a \div T \times f \div C_{pr} \times 1000$$

[Note]

 ΔA_{400} : The change OD value of sample well. ($\Delta A_{400} = A_{sample} - A_{control}$).

T: The time of reaction, 10 min.

f: Dilution factor of sample before test.

C_{pr}: The concentration of protein in sample, gprot/L.

1000: 1 mmol/L=1000 μmol/L.

Appendix I Performance Characteristics

1. Parameter:

Intra-assay Precision

Three human serum samples were assayed in replicates of 20 to determine precision within an assay (CV = Coefficient of Variation).

Parameters	Parameters Sample 1		Sample 3		
Mean (U/L)	0.50	2.50 5.0			
% CV 5.0		3.2	4.4		

Inter-assay Precision

Three human serum samples were assayed 20 times in duplicate by three operators to determine precision between assays.

Parameters	Parameters Sample 1		Sample 3		
Mean (U/L)	0.50	2.50	5.00		
% CV 6.8		4.6	4.8		

Recovery

Take three samples of high concentration, middle concentration and low concentration to test the samples of each concentration for 6 times parallelly to get the average recovery rate of 96%.

	Standard 1	Standard 2	Standard 3
Expected Conc. (mmol/L)	0.12	0.25	0.38
Observed Conc. (mmol/L)	0.1	0.2	0.4
recovery rate(%)	92	98	98

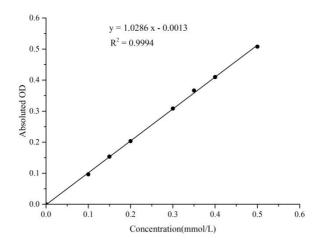
Sensitivity

The analytical sensitivity of the assay is 0.76 U/L. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 20 times, and calculating the corresponding concentration

2. Standard curve:

As the OD value of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique or temperature effects), so the standard curve and data are provided as below for reference only:

Concentration (mmol/L)	0	0.1	0.15	0.2	0.3	0.35	0.4	0.5
OD value	0.064	0.161	0.218	0.267	0.372	0.435	0.471	0.573
	0.065	0.161	0.218	0.269	0.374	0.427	0.478	0.572
Average OD	0.065	0.161	0.218	0.268	0.373	0.431	0.475	0.573
Absoluted OD	0.000	0.097	0.154	0.204	0.309	0.367	0.410	0.508



Appendix Π Example Analysis

Example analysis:

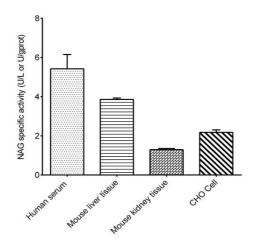
For 10% mouse liver tissue homogenate, dilute for 4 times, take 20 μ L of diluted sample and carry the assay according to the operation table. The results are as follows:

standard curve: y = 1.0286x - 0.0013. The OD value of the control well is 0.089, the OD value of the sample well is 0.178, $\Delta A400$ =Asample - Acontrol = 0.178 - 0.089 = 0.089. The concentration of protein in sample is 9.00 gprot/L, and the calculation result is:

NAG activity(U/gprot) =
$$(0.089 + 0.0013) \div 1.0286 \div 10 \times 4 \div 9.00 \times 1000$$

= 3.90 U/gprot

Detect human serum, 10% mouse liver tissue homogenate (the concentration of protein is 9.00 gprot/L, dilute for 4 times), 10% mouse kidney tissue homogenate (the concentration of protein is 9.84 gprot/L, dilute for 4 times), and CHO cell (the concentration of protein is 0.14 gprot/L) according to the protocol, the result is as follows:



Statement

- 1. This assay kit is for Research Use Only. We will not response for any arising problems or legal responsibilities causing by using the kit for clinical diagnosis or other purpose.
- Please read the instructions carefully and adjust the instruments before the experiments. Please follow the instructions strictly during the experiments.
- 3. Protection methods must be taken by wearing lab coat and latex gloves.
- 4. If the concentration of substance is not within the detection range exactly, an extra dilution or concentration should be taken for the sample.
- 5. It is recommended to take a pre-test if your sample is not listed in the instruction book.
- 6. The experimental results are closely related to the situation of reagents, operations, environment and so on. Elabscience will guarantee the quality of the kits only, and NOT be responsible for the sample consumption caused by using the assay kits. It is better to calculate the possible usage of sample and reserve sufficient samples before use.