

(FOR RESEARCH USE ONLY. DO NOT USE IT IN CLINICAL DIAGNOSIS !)

**Catalog No: E-BC-K153-M**

**Specification: 48T(24 samples)/96T(48 samples)**

**Measuring instrument: Microplate reader (330-350 nm)**

**Detection range: 3.55-74.21 U/L**

**Elabscience<sup>®</sup> Mitochondrial Complex V  
(F<sub>0</sub>F<sub>1</sub>-ATPase/ATP Synthase) Activity Assay Kit**

This manual must be read attentively and completely before using this product.  
If you have any problem, please contact our Technical Service Center for help:

Toll-free: 1-888-852-8623

Tell: 1-832-243-6086

Fax: 1-832-243-6017

Email: [techsupport@elabscience.com](mailto:techsupport@elabscience.com)

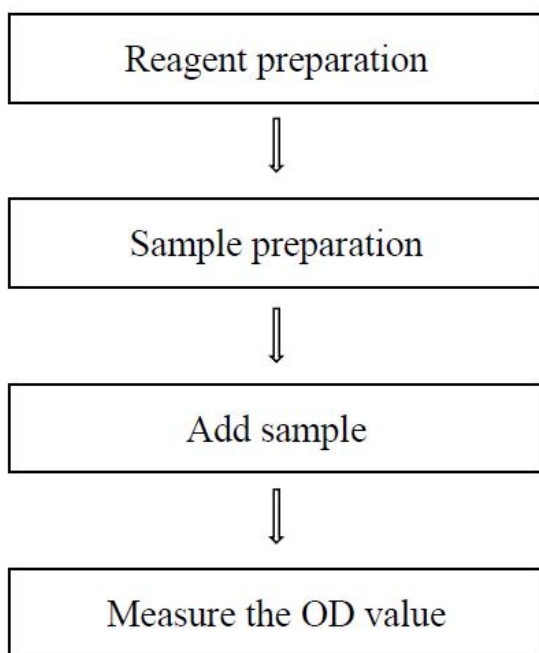
Website: [www.elabscience.com](http://www.elabscience.com)

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

## Table of contents

<b>Assay summary .....</b>	<b>3</b>
<b>Intended use .....</b>	<b>4</b>
<b>Detection principle .....</b>	<b>4</b>
<b>Kit components &amp; storage .....</b>	<b>4</b>
<b>Materials prepared by users .....</b>	<b>5</b>
<b>Reagent preparation .....</b>	<b>5</b>
<b>The key points of the assay .....</b>	<b>7</b>
<b>Calculation .....</b>	<b>9</b>
<b>Appendix I Performance Characteristics .....</b>	<b>10</b>
<b>Appendix II Example Analysis .....</b>	<b>11</b>
<b>Appendix III Publications .....</b>	<b>12</b>
<b>Statement .....</b>	<b>13</b>

## Assay summary



## Intended use

This kit can measure mitochondrial complex V ( $F_0F_1$ -ATPase/ATP Synthase) in animal tissue samples.

## Detection principle

Mitochondrial complex V is also known as  $F_0F_1$ -ATP synthase. ATP is hydrolyzed by  $F_0F_1$ -ATP synthase to produce ADP, and ADP converts NADH into oxidized coenzyme I ( $NAD^+$ ) after enzyme conversion reaction. Therefore, the activity of mitochondrial complex V can be quantified by measure the change OD value at 340 nm.

## Kit components & storage

Item	Component	Size 1(48 T)	Size 2(96 T)	Storage
Reagent 1	Extraction Solution A	50 mL × 1 vial	50 mL × 2 vials	-20°C, 12 months
Reagent 2	Extraction Solution B	25 mL × 1 vial	50 mL × 1 vial	-20°C, 12 months
Reagent 3	Protease Inhibitor	0.8 mL × 1 vial	0.8 mL × 2 vials	-20°C, 12 months, shading light
Reagent 4	Buffer Solution	15 mL × 1 vial	30 mL × 1 vial	-20°C, 12 months
Reagent 5	Substrate A	Liquid × 1 vial	Liquid × 2 vials	-20°C, 12 months, shading light
Reagent 6	Substrate B	Powder × 1 vial	Powder × 2 vials	-20°C, 12 months, shading light
Reagent 7	Substrate C	Powder × 1 vial	Powder × 2 vials	-20°C, 12 months, shading light
Reagent 8	Substrate D	Powder × 2 vials	Powder × 4 vials	-20°C, 12 months, shading light
Reagent 9	Inhibitor	0.05 mL × 1 vial	0.1 mL × 1 vial	-20°C, 12 months, shading light
	UV Microplate	96 wells		No requirement
	Plate Sealer	2 pieces		

Note: The reagents must be stored strictly according to the preservation conditions

in the above table. The reagents in different kits cannot be mixed with each other. For a small volume of reagents, please centrifuge before use, so as not to obtain sufficient amount of reagents.

## **Materials prepared by users**

### **Instruments:**

Centrifuge, 37°C incubator, Microplate reader (330-350 nm, optimum wavelength: 340 nm)

## **Reagent preparation**

① Equilibrate all the reagents to room temperature before use.

② The preparation of Substrate B working solution:

Dissolve one vial of substrate B with 300  $\mu\text{L}$  of double distilled water, mix well. Aliquot and store at -20°C for 7 days protected from light.

③ The preparation of Substrate C working solution:

Dissolve one vial of substrate C with 300  $\mu\text{L}$  of double distilled water, mix well. Aliquot and store at -20°C for 7 days protected from light.

④ The preparation of reaction working solution:

Before testing, please prepare sufficient reaction working solution according to the test wells. For example, prepare 525  $\mu\text{L}$  of reaction working solution (mix well 500  $\mu\text{L}$  of buffer solution, 5  $\mu\text{L}$  of Substrate A, 10  $\mu\text{L}$  of Substrate B working solution, 10  $\mu\text{L}$  of Substrate C working solution). The prepared solution can be stored at 2-8°C for 8 h protected from light.

⑤ The preparation of enzyme working solution:

Dissolve one vial of substrate D with 0.6 mL of double distilled water, mix well. Store at 2-8°C for 3 days protected from light.

⑥ The preparation of specific working solution:

Before testing, please prepare sufficient specific working solution according to the test wells. For example, prepare 505  $\mu\text{L}$  of specific working solution (mix well 5  $\mu\text{L}$  of inhibitor and 500  $\mu\text{L}$  of double distilled water). The specific working solution should be prepared on spot. The remaining inhibitor can be stored at  $-20^{\circ}\text{C}$  for 1 week.

## **Sample preparation**

### **① Sample preparation**

#### **Tissue sample:**

- ① Harvest the amount of tissue needed for each assay (initial recommendation 100 mg).
- ② Wash tissue in cold PBS (0.01 M, pH 7.4).
- ③ Homogenize 100 mg tissue in 900  $\mu\text{L}$  extraction solution A with a dounce homogenizer at  $4^{\circ}\text{C}$ .
- ④ Centrifuge at  $600\times g$  for 5 min, discard the precipitate and take the supernatant.
- ⑤ Then centrifuge at  $15000\times g$  for 10 min at  $4^{\circ}\text{C}$ , discard the supernatant and take the precipitate.
- ⑥ The precipitate was mixed with 200  $\mu\text{L}$  of extraction solution B and 10  $\mu\text{L}$  of protease inhibitor, sonicated for 5 min, centrifuged at  $15000\times g$  at  $4^{\circ}\text{C}$  for 10 min. Then take the supernatant for detection.
- ⑦ Meanwhile, determine the protein concentration of supernatant (E-BC-K318-M).

## ② Dilution of sample

The recommended dilution factor for different samples is as follows (for reference only):

Sample type	Dilution factor
10% Mouse liver tissue homogenate	2-6
10% Rat liver tissue homogenate	1-4
10% Mouse kidney tissue homogenate	2-6
10% Rat muscle tissue homogenate	1-3

Note: The diluent is extraction solution B. For the dilution of other sample types, please do pretest to confirm the dilution factor.

## The key points of the assay

- ① During reagent preparation, it is necessary to ensure that the powder is completely dissolved in the reaction working solution after preparation.
- ② The detection is started at about 10 s after adding reaction working solution.
- ③ For sample detection, if the  $A_1$  of the sample well and the control well is lower than 0.7, or the change OD value ( $\Delta A$ ) of the sample well and the control well for 4 min exceeds 0.3, the sample should be diluted.

## Operating steps

- ① Control well: Add 10  $\mu\text{L}$  of specific working solution to the corresponding wells.  
Sample well: Add 10  $\mu\text{L}$  of double distilled water to the corresponding wells.
- ② Add 20  $\mu\text{L}$  of sample to each well.
- ③ Mix fully with microplate reader for 3 s. And incubate at  $37^{\circ}\text{C}$  for 4 min
- ④ Add 20  $\mu\text{L}$  of enzyme working solution to each well.
- ⑤ Add 180  $\mu\text{L}$  of reaction working solution to each well.
- ⑥ Measure the OD value of each well at 340 nm with microplate reader, recorded as  $A_1$ . After 4 min, measure the OD value of each well at 340 nm with microplate reader, recorded as  $A_2$ .  $\Delta A = A_1 - A_2$ . (It is recommended to follow the notes below for measurement)

**Note:** The sample wells measure the total enzyme activity, and the control wells measure the non-specific enzyme activity. After adding the reaction working solution, record the OD value once every minute for 4 min, observe the change of OD value within 4 min to ensure whether is a constant rate of decline. Otherwise, the sample needs to be diluted. When calculating, take initial OD value  $A_1$ ,  $A_2$  OD value after 4 min.



## Calculation

**The sample:**

**For tissue samples:**

**Definition:** The amount of mitochondrial complex V in 1 g mitochondrial protein per 1 minute that catalyze decomposition of 1  $\mu\text{mol}$  NADH at 37°C is defined as 1 unit.

mitochondrial complex V activity =  
(U/gprot)

$$\frac{\Delta A_{\text{sample}} - \Delta A_{\text{control}}}{6220 \times 0.65} \times 0.23 \div t \div 0.02 \div C_{\text{pr}} \times f \times 10^6$$

### [Note]

$\Delta A_{\text{sample}}$ : The change OD value of sample well,  $A_1 - A_2$ .

$\Delta A_{\text{control}}$ : The change OD value of control well,  $A_1 - A_2$ .

6220: Molar absorption coefficient,  $\text{L}/(\mu\text{mol} \cdot \text{cm})$ .

0.65: Optical path, cm

0.23: The volume of the reaction system, mL.

0.02: The volume of the sample, mL.

T: The time of reaction, 4 min.

f: Dilution factor of sample before test.

$C_{\text{pr}}$ : The concentration of protein in sample, gprot/L.

$10^6$ :  $1 \text{ mol} = 10^6 \mu\text{mol}$ .

## Appendix I Performance Characteristics

### 1. Parameter:

#### Intra-assay Precision

Three mouse kidney tissue samples were assayed in replicates of 20 to determine precision within an assay. (CV = Coefficient of Variation)

Parameters	Sample 1	Sample 2	Sample 3
Mean (U/L)	21.50	38.50	65.60
%CV	5.8	5.6	5.4

#### Inter-assay Precision

Three mouse kidney tissue samples were assayed 20 times in duplicate by three operators to determine precision between assays.

Parameters	Sample 1	Sample 2	Sample 3
Mean (U/L)	21.50	38.50	65.60
%CV	6.4	6.3	6.8

#### Recovery

Take three samples of high concentration, middle concentration and low concentration to test the samples of each concentration for 6 times parallelly to get the average recovery rate of 101%.

	Sample 1	Sample 2	Sample 3
Expected Conc. (U/L)	25	32.4	70.2
Observed Conc. (U/L)	25.8	32.1	70.9
Recovery rate (%)	103	99	101

#### Sensitivity

The analytical sensitivity of the assay is 3.55 U/L. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 20 times, and calculating the corresponding concentration.

## Appendix II Example Analysis

### Example analysis:

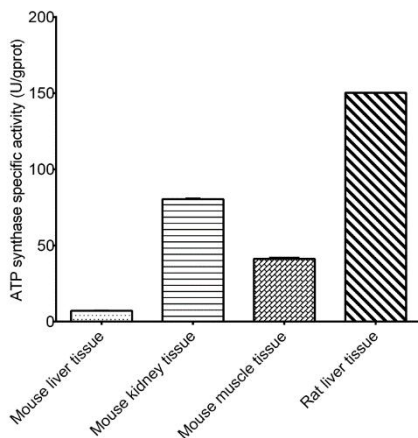
For 10% mouse liver tissue mitochondria supernatant, dilute for 4 times, carry the assay according to the operation steps. The results are as follows:

The  $A_1$  of the sample well is 0.900, the  $A_2$  of the sample well is 0.426,  $\Delta A_{\text{sample}} = 0.900 - 0.426 = 0.474$ . The  $A_1$  of control well is 0.914, the  $A_2$  of control well is 0.471,  $\Delta A_{\text{control}} = 0.914 - 0.471 = 0.443$ , the concentration of mitochondria protein in sample is 12.14 gprot/L, and the calculation result is:

mitochondrial complex V activity  
(U/gprot)

$$= \frac{0.474 - 0.443}{6220 \times 0.65} \times 0.23 \div 4 \div 0.02 \div 12.14 \times 4 \times 10^6 = 7.26 \text{ U/gprot}$$

Detect 10% mouse liver tissue homogenate (the concentration of mitochondria protein is 12.14 gprot/L, dilute for 4 times), 10% mouse kidney tissue homogenate (the concentration of mitochondria protein is 12.33 gprot/L, dilute for 4 times), 10% mouse muscle tissue homogenate (the concentration of mitochondria protein is 1.86 gprot/L, dilute for 4 times) and 10% rat liver tissue homogenate (the concentration of mitochondria protein is 7.50 gprot/L, dilute for 4 times) according to the protocol, the result is as follows:



### Appendix III Publications

1. Tian Y , Hong X , Xie Y ,et al.17  $\beta$  -Estradiol (E 2) Upregulates the ER  $\alpha$  /SIRT1/PGC-1  $\alpha$  Signaling Pathway and Protects Mitochondrial Function to Prevent Bilateral Oophorectomy (OVX)-Induced Nonalcoholic Fatty Liver Disease (NAFLD)[J].Antioxidants, 2023, 12(12).DOI:10.3390/antiox12122100.
2. Wang Y , Gao J , Fan B ,et al.Different levels of autophagy induced by transient serum starvation regulate metabolism and differentiation of porcine skeletal muscle satellite cells[J].Scientific reports, 13(1):13153[2025-03-04].DOI:10.1038/s41598-023-40350-y.
3. Gu W, Cong X, Pei Y, et al. Impaired Mitochondrial Energy Metabolism Regulated by p70S6K: A Putative Pathological Feature in Alzheimer' s Disease[J]. Metabolites, 2024, 14(7): 369.
4. Xiao P , Wu S , Wang Z ,et al.Biototoxicity of paraquat to lung cells mediated by endoplasmic reticulum-mitochondria interaction[J].Journal of Molecular Histology, 2024, 55(6):1063-1077.DOI:10.1007/s10735-024-10249-7.
5. Chen L , Chen S , Bai Y ,et al.Electroacupuncture improves cognitive impairment after ischemic stroke based on regulation of mitochondrial dynamics through SIRT1/ PGC-1  $\alpha$  pathway[J].Brain research, 2024:1844.DOI:10.1016/j.brainres.2024.149139.

## Statement

1. This assay kit is for Research Use Only. We will not response for any arising problems or legal responsibilities causing by using the kit for clinical diagnosis or other purpose.
2. Please read the instructions carefully and adjust the instruments before the experiments. Please follow the instructions strictly during the experiments.
3. Protection methods must be taken by wearing lab coat and latex gloves.
4. If the concentration of substance is not within the detection range exactly, an extra dilution or concentration should be taken for the sample.
5. It is recommended to take a pre-test if your sample is not listed in the instruction book.
6. The experimental results are closely related to the situation of reagents, operations, environment and so on. Elabscience will guarantee the quality of the kits only, and NOT be responsible for the sample consumption caused by using the assay kits. It is better to calculate the possible usage of sample and reserve sufficient samples before use.





