

**FB1 (Fumonisin B1) ELISA Kit**

Catalog No: E-TO-E020

96T/96T\*3

<b>Version Number:</b>	V1.5
<b>Replace version:</b>	V1.4
<b>Revision Date:</b>	2026.05.29

This manual must be read attentively and completely before using this product.

If you have any problems, please contact our Technical Service Center for help.

Toll-free: 1-888-852-8623 Tel: 1-832-243-6086 Fax: 1-832-243-6017

Email: [techsupport@elabscience.com](mailto:techsupport@elabscience.com)

Website: [www.elabscience.com](http://www.elabscience.com)

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

## Test principle

This kit uses Competitive-ELISA as the method for the quantitative detection. It can detect Fumonisin B1 (FB1) in samples, such as corn, feed, etc. This kit is composed of ELISA Microtiter plate, HRP conjugate, antibody working solution, standard and other supplementary reagents. The microtiter plate in this kit has been pre-coated with coupled antigen. During the reaction, FB1 in the samples or standard competes with coupled antigen on the solid phase supporter for sites of anti-FB1 antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each microtiter plate well, and substrate reagent is added for color development. There is a negative correlation between the OD value of samples and the concentration of FB1. The concentration of FB1 in the samples can be calculated by comparing the OD of the samples to the standard curve.

## Technical indicator

**Reaction mode** (Incubation time and temperature): 25±2°C; 30 min, 15 min.

**Detection limit:** Corn, feed---50 ppb; Edible oil---20 ppb.

**Cross-reactivity:** Fumonisin B1 (FB1) ---100%

**Sample recovery rate:** Feed---95%±15%; Corn---100%±15%; Edible oil---85%±15%.

## Kits components

Item	Specifications
ELISA Microtiter plate	96 wells
Standard Liquid	1/1*3 mL each (ppb=ng/mL=ng/g) (0 ppb, 1 ppb, 3 ppb, 9 ppb, 27 ppb, 81 ppb)
HRP Conjugate	5.5/5.5*3 mL
Antibody Working Solution	5.5/5.5*3 mL
Substrate Reagent A	6/6*3 mL
Substrate Reagent B	6/6*3 mL
Stop Solution	6/6*3 mL
10×Reconstitution Buffer	50/50*3 mL
20×Concentrated Wash Buffer	40/40*3 mL
Plate Sealer	1/1*3 piece
Sealed Bag	1/1*3 piece
Manual	1 copy

Note: All reagent bottle caps must be tightened to prevent evaporation and microbial pollution.

---

**Other materials required but not supplied**

**Instrument:** Microplate reader, Printer, Homogenizer, Vortex mixer, Centrifuge, Graduated pipette, Balance (sensitivity 0.01 g).

**High-precision transferpeltor:** Single channel (20-200 µL, 100-1000 µL), Multichannel (300 µL).

**Reagents:** Methanol, N-hexane.

**Notes**

1. The overall OD value will be lower when reagents have not been brought to room temperature before use or room temperature is below 25±2°C.
2. If the wells turn dry during the washing procedure, it will lead to bad linear standard curve and poor repeatability. Operate the next step immediately after wash.
3. Mix thoroughly and wash the plate completely. The consistency of wash procedure can strongly affect the reproducibility of this ELISA kit.
4. FOR RESEARCH USE ONLY. ELISA Microtiter plate should be covered by plate sealer. Avoid the kit to strong light.
5. **Each reagent is optimized for use in the E-TO-E020. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other E-TO-E020 with different lot numbers.**
6. Substrate Reagent should be abandoned if it turns blue color. When OD value of standard (concentration: 0) < 0.5 unit (A450nm < 0.5), it indicates the reagent may be deteriorated.
7. Stop solution is caustic, avoid contact with skin and eyes.
8. As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.
9. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
10. **For mentioned sample fast and efficient extraction methods are included in the kit description. Please consult technical support for the applicability if other sample need to be tested.**
11. The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS, etc. can be used for quantitative confirmation.

**Storage and expiry date**

Store the kit at 2-8°C. Do not freeze any test kit components.

Return any unused microwells to their original foil bag and reseal them together with the desiccant provided and further store at 2-8°C. After opening, the kit is stable for up to 1 month.

**Expiry date:** expiration date is on the packing box.

---

## Experimental preparation

Restore all reagents and samples to room temperature before use.

Open the micro-plate reader in advance, preheat the instrument, and set the testing parameters.

### 1. Sample pretreatment Notice:

Experimental apparatus should be clean, and use disposable pipette tips to avoid cross-contamination during the experiment.

### 2. Solution preparation

*Please prepare solution according to the number of samples. Don't use up all components at once!*

#### Solution 1: Reconstitution Buffer

Dilute the **10×Reconstitution Buffer** with deionized water (10×Reconstitution Buffer (V): Deionized water (V) =1:9). Store at 4°C for a month.

#### Solution 2: Wash Buffer

Dilute the **20×Concentrated Wash Buffer** with deionized water. (20×Concentrated Wash Buffer (V): Deionized water (V) =1:19)

#### Solution 3: 70% Methanol

Dilute **Methanol** with deionized water. (Methanol (V): Deionized water (V) =7:3)

### 3. Sample pretreatment procedure

#### 3.1 Pretreatment of corn, feed sample:

- (1) Homogenize the representative sample with a homogenizer and mix fully.
- (2) Weigh  $1\pm 0.05$  g of crushed homogenate into the 50 mL centrifuge tube, add 5 mL of deionized water, vortex for 5 min. Centrifuge at 4000 r/min for 10 min at room temperature.
- (3) Take 0.1 mL of supernatant, add 0.9 mL of **Reconstitution Buffer** (Solution 1), vortex for 30 s.
- (4) Take 50  $\mu$ L for analysis.

**Note: Sample dilution factor: 50, detection limit: 50 ppb**

*(For samples with extremely high levels of toxins, the Reconstitution Buffer (Solution 1) can be used to further dilute before testing. For example, take 0.1 mL of the mixed solution from step 3 of 3.1, add 0.9 mL of Reconstitution Buffer (Solution 1) and mix fully. The final sample has a dilution ratio of 500 and detection limit of 500 ppb. Note: Due to the uneven distribution of toxins in the sample, it is necessary to take multiple samples and mix thoroughly before taking 1 g for testing.)*

#### 3.2 Pretreatment of edible oil sample:

- (1) Take 5 mL of sample into the 50 mL centrifuge tube, add 8 mL of **N-hexane** and **70% Methanol** (Solution 3), vortex for 5 min. Centrifuge at 4000 r/min for 10 min at room temperature.
- (2) Take 0.1 mL of supernatant, add 0.9 mL of **Reconstitution Buffer** (Solution 1), vortex for 30 s.
- (3) Take 50  $\mu$ L for analysis.

**Note: Sample dilution factor: 20, detection limit: 20 ppb**

---

## Assay procedure

Restore all reagents and samples to room temperature ( $25\pm 2^{\circ}\text{C}$ ) before use. All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming. The unused ELISA Microtiter plate should be sealed as soon as possible and stored at  $2-8^{\circ}\text{C}$ .

1. **Number:** number the sample and standard in order (multiple well), and keep a record of standard wells and sample wells. **Standard and Samples need test in duplicate.**
2. **Add sample:** add  $50\ \mu\text{L}$  of **Standard or Sample** per well, then add  $50\ \mu\text{L}$  of **HRP Conjugate** to each well, then add  $50\ \mu\text{L}$  of **Antibody Working Solution**, cover the plate with plate sealer, oscillate for 5 s gently to mix thoroughly. Incubate at  $25\pm 2^{\circ}\text{C}$  for 30 min in shading light.
3. **Wash:** uncover the sealer carefully, remove the liquid. Immediately add  $350\ \mu\text{L}$  of **Wash Buffer** (Solution 2) to each well and wash. Repeat wash procedure for 5 times, 30 s intervals/time. Invert the plate and pat it against thick clean absorbent paper (If bubbles exist in the wells, clean tips can be used to prick them).
4. **Color Development:** add  $50\ \mu\text{L}$  of **Substrate Reagent A** to each well, and then add  $50\ \mu\text{L}$  of **Substrate Reagent B**. Gently oscillate for 5 s to mix thoroughly. Incubate at  $25\pm 2^{\circ}\text{C}$  for 15 min in shading light (The reaction time can be extended according to the actual color change).
5. **Stop reaction:** add  $50\ \mu\text{L}$  of **Stop Solution** to each well, oscillate gently to mix thoroughly.
6. **OD Measurement:** determine the optical density (OD value) of each well at 450 nm (reference wavelength 630 nm) with a microplate reader. This step should be finished in 10 min after stop reaction.

## Result analysis

### 1. Absorbance (%) = $A/A_0 \times 100\%$

A: Average absorbance of standard or samples

$A_0$ : Average absorbance of 0 ppb Standard

### 2. Drawing and calculation of standard curve

Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add average absorbance value of sample to standard curve to get corresponding concentration. **If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.**

For this kit, it is more convenient to use professional analysis form for accurate and fast analysis on a large number of samples.

### Fumonisin B1 (E-TO-E020) Standard Curve

