EasySort™ Mouse CD8⁺T Cell Isolation Kit

Cat. No: MIM003N **Size: 10Assays/100Assays/200Assays**

Component	Component Name	10 Assays	100 Assays	200 Assays	Storage
MIM003NA	EasySort™ Mouse CD8⁺T Beads Streptavidin 1.0-N	150 μL	1500 μL	1500 μL×2	2~8°C
MIM003NB	EasySort™ Mouse CD8⁺T Cell Isolation Cocktail	50 μL	500 μL	500 μL×2	2~8°C
	Manual	1 copy			

Storage

Store at 2-8°C with shading light for 1 year. Avoid freezing and thawing.

Detection Principle

The EasySort™ Mouse CD8⁺T cell Isolation Kit is a product that enables rapid and simple isolation of high-purity mouse CD8⁺T cells. This kit uses a negative selection method and is suitable for isolating CD8⁺T cells from mouse spleen samples. Different biotinylated monoclonal antibodies are used to label non-target cells (non-mouse CD8⁺ T cells). Subsequently, streptavidin-conjugated magnetic beads are employed to deplete these non-target cells, thereby obtaining highly purified mouse CD8⁺T cells. The isolated mouse CD8⁺T cells are free of any antibodies and magnetic bead labels, remain in an unstimulated, naïve state, and are ready for direct use in downstream applications.

The EasySort™ Mouse CD8⁺T cell Isolation Kit has been tested by magnetic cell separation followed by flow cytometric analysis of cells from mouse spleen tissues. An assay is defined as 5 μL antibody and 15 μL beads to be used to isolate $1x10^7$ cells..

Reagents and Materials Not Supplied

1. **Reagents:**

   PBS, fetal bovine serum (FBS), EDTA

2. **Materials:**

   Disposable sterile syringe, 70 μm mesh nylon strainer, ophthalmic scissors, ophthalmic forceps, 1.5 mL/2 mL EP tube, 15 mL centrifuge tube, flow tube

3. **Instrument:**

   Optical microscope, horizontal centrifuge, magnetic rack

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Experimental Operation

NOTE: The following operations must be performed under sterile conditions

- **Isolation buffer preparation**

  Add fetal bovine serum (final concentration of 2%) and EDTA (final concentration of 2 mM) to PBS buffer and filter the prepared buffer with 0.22 μm filter.

  NOTE: Sealed store the prepared buffer at 4°C and use within 1 week. In addition, 2% fetal bovine serum can be replaced by 0.5% bovine serum albumin (BSA).

- **Mouse spleen single cell suspension preparation**

a. Take the fresh mouse spleen to avoid excessive connective tissue attached.

b. Grind the spleen through a 70 μm mesh nylon strainer, rinse the cell sieve with pre-cooled PBS, and collect the cell suspension in a 15 mL centrifuge tube and centrifuge at 300 g for 5 min.

c. Discard the supernatant, resuspend the splenocytes with isolation buffer, and filter the cells through a 70 μm mesh nylon strainer, then count the cells. Adjust the cell density to 2×10^8 cells/mL.

   Note: Generally, approximately $2\text{-}4\times10^8$ splenocytes can be obtained from each mouse. After preparing a single-cell suspension from mouse spleen perform the cell isolation experiment within 1-2 hours, as a longer interval will affect the final isolated cell purity and cell viability.

- **Cell Sorting**

a. Prepare 50 μL of cell suspension (about 1×10^7 cells), add 5 μL Mouse $CD8^+$T Cell Isolation Cocktail, mix fully and incubate for 5 min at room temperature.

   Note: Please make sure the cells are single-cell suspension.

b. Add isolation buffer to a final volume of 2 mL, centrifuge at 300 g for 5 min. Discard the supernatant, and then resuspend the cells with 50 μL isolation buffer.

   Note:

   - If the total volume of the cell suspension exceeds 1 mL, the volume of the added isolation buffer shall be no less than the total volume of the cell suspension. For example, if the total volume of the cell suspension is 1.5 mL, the volume of the isolation buffer added shall be ≥ 1.5 mL.
   - To maintain consistent cell density, the volume of cell isolation buffer for cell resuspension shall be identical to that of the input cell suspension. In the protocol example, if 50 μL of cell suspension is used as the starting input, cells should be resuspended with an equal volume of 50 μL cell isolation buffer.

c. Wash Beads Streptavidin 1.0-N: Place a clean flow cytometry tube or a centrifuge tube compatible with the magnetic rack into a tube rack. Pipette 1 mL of isolation buffer into the tube, vortex beads for 20 seconds, then add 15 μL of magnetic beads directly into the aforementioned 1 mL of isolation buffer. Mix by pipetting up and down 6-8 times. Place the flow cytometry tube or centrifuge tube on a magnetic rack (provided by the user) and magnetically separate at room temperature for 5 min. At this point, the magnetic beads are attracted to the tube wall. Keep the tube on the magnetic rack, discard the supernatant, and then remove the tube from the magnetic rack.

   Note： If the total volume of magnetic beads to be washed is greater than 1 mL, use a 1:1 volume ratio of isolation buffer to beads during the washing step.

d. Resuspend the magnetic beads using the cell suspension from step b): Aspirate the cell suspension and pipette the beads off the tube wall to the bottom of the tube (Note: avoid

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generating bubbles). Mix by pipetting up and down 6-8 times, then incubate at room temperature for 5 min.

Note:

- If more than 1×10^7 cells are to be sorted, increase the amount of Mouse CD8$^+$T Cell Isolation Cocktail and Mouse CD8$^+$T Beads Streptavidin 1.0-N proportionally while ensuring the cell density remains 2×10^8 cells/mL. If fewer than 1×10^7 cells are to be sorted, resuspend the cells with 50 µL sorting buffer, add 5 µL Mouse CD8$^+$T Cell Isolation Cocktail and 15 µL washed Mouse CD8$^+$T Beads Streptavidin 1.0-N.
- The 5 mL flow tube is suitable for isolation of cell suspension ≤ 1 mL (2×10^8 cells). 10 mL or 15 mL centrifuge tube is suitable for isolation of cell suspension ≤ 4 mL (8×10^8 cells).

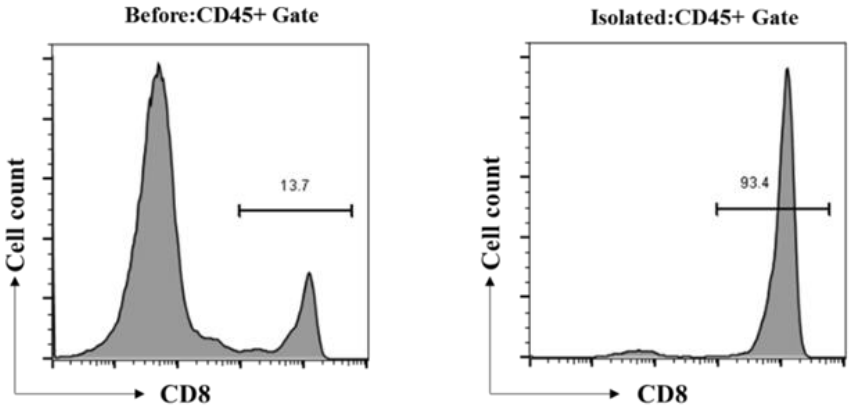
e. Add isolation buffer to a final volume of 2.5 mL (If the volume of the cell suspension for isolation is >1 mL, resuspend in an equal volume of isolation buffer), mix fully with a pipette by blowing up and down for 6-8 times until no particles of magnetic beads are visible. Put the tube on a 5 mL magnetic rack (self-provided) and stand for 5 min.

Note: Please mix the liquid thoroughly to avoid the magnetic beads clumping and affecting the isolation efficiency.

f. Transfer the cell suspension to a clean centrifuge tube, this is the CD8$^+$T cells obtained from the first isolation. Add isolation buffer to a final volume of 2.5 mL into the flow tube, mix with a pipette by blowing up and down for 7-8 times until no particles of magnetic beads are visible. Put the tube on a 5 mL magnetic rack (self-provided) and stand for 5 min.

g. Transfer the cell suspension to the centrifuge tube in step f, mix the cell suspension obtained from step e and f, centrifuge at 300 g for 5 min. Discard the supernatant, resuspend the cells with isolation buffer required for the subsequent experiments.

Typical data



CD8$^+$T cells were isolated from the spleen cells of C57BL/6 mice, and were stained with Elab Fluor® Violet 450 Anti-Mouse CD45 Antibody (clone 30-F11) and PE Anti-Mouse CD8a Antibody (clone 53-6.7). The purities of the start and final isolated fractions were 13.7% and 93.4%, respectively.

Cautions

1. This kit is for research use only.

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2. Please take safety precautions and follow the procedures of laboratory reagent operation.
3. Avoid freezing and thawing during the use and storage of the beads.
4. Single-cell suspension for cell isolation shall be filtered through a cell strainer to remove cell clumps and tissue debris, preventing cell aggregation from compromising isolation purity.
5. Perform isolation immediately after preparing the cell suspension, as cell viability will decrease with longer storage time.
6. When adding the antibody cocktail and aspirating the magnetic beads for washing, pipette them directly to the bottom of the tube to avoid adhesion to the wall, which would result in loss of components.
7. In order to ensure the activity of the cells, the whole process of the experiment should be completed on ice as much as possible, except for the incubation at room temperature.
8. It is recommended to use low adsorption pipette tips and centrifuge tubes to avoid the loss of magnetic beads and antibodies due to adsorption.
9. The kit should be used in conjunction with a magnetic rack.
10. Sample type, sample preparation and experimental operation have an important impact on the final isolated cell purity.

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