EasySort™ Human CD8+ T Cell Isolation Kit

Cat. No: MIH003N **Size: 10 Assays/100 Assays/200 Assays**

Component	Component Name	10 Assays	100 Assays	200 Assays	Storage
MIH003NA	EasySort™ Human CD8+ T Beads Streptavidin 1.0-N	240 μL	1.2 mL×2	1.2 mL×4	2-8°C
MIH003NB	EasySort™ Human CD8+ T Cell Isolation Cocktail	300 μL	1.5 mL×2	1.5 mL×4	2-8°C
	Manual	1 copy			

Storage

Store at 2-8°C with shading light for 1 year. Avoid freezing and thawing.

Description

The EasySort™ Human CD8+T Cell Isolation Kit is a product that enables rapid and simple isolation of high-purity human CD8+T cells. This kit employs a negative selection method and is suitable for isolating CD8+T cells from either fresh or cryopreserved human PBMC samples. Different biotinylated monoclonal antibodies are used to label non-target cells (non-human CD8+ T cells). Subsequently, streptavidin-conjugated magnetic beads are employed to deplete these non-target cells, thereby obtaining highly purified human CD8+ T cells. The isolated human CD8+T cells carry no antibody or magnetic bead labels, remain in an undisturbed, naïve state, and are ready for downstream applications.

The EasySort™ Human CD8+T Cell Isolation Kit has been validated for magnetic separation using fresh or cryopreserved human PBMC samples, and the isolated cells were analyzed and characterized by flow cytometry. The kit's single experiment protocol requires 24 μL of magnetic beads and 30 μL of antibody cocktail, sufficient for isolating up to 1 × 10^7 cells.

Reagents and Materials Not Supplied

1. **Reagents:**

   PBS, fetal bovine serum (FBS), EDTA, Human peripheral blood mononuclear cells separation solution、DNase I

2. **Materials:**

   70 μm mesh nylon strainer, 1.5 mL/2 mL EP tube, 15 mL/50 mL centrifuge tube, flow tube

3. **Instrument:**

   Optical microscope, horizontal centrifuge, magnetic rack

Experimental Operation

Toll-free: 1-888-852-8623 Tel: 1-832-243-6086 Fax: 1-832-243-6017
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NOTE: The following operations must be performed under sterile conditions

➢ Isolation buffer preparation

Add fetal bovine serum (final concentration of 2%) and EDTA (final concentration of 2 mM) to PBS buffer and filter the prepared buffer with 0.22 μm filter.

NOTE: Sealed store the prepared buffer at 4°C and use within 1 week. In addition, 2% fetal bovine serum can be replaced by 0.5% bovine serum albumin (BSA) .

➢ Sample Preparation and Processing

1. Fresh human PBMC: PBMC sample is obtained from fresh human whole blood by density gradient centrifugation. Wash PBMC twice with isolation buffer, centrifuge at 300 g for 5 minutes, filter the PBMC through a 70 μm mesh nylon strainerand and adjust the cell density to 1×10^8 cells/mL for cell isolation.

   Note: The best separation effect can be achieved when the freshly collected human blood is separated within 1 hour. Approximately 1×10^7 PBMC can be obtained from 10 mL of human blood.

2. Frozen PBMC: The frozen PBMC should be incubated with DNase I solution (PBS) at a concentration of 100 μg/mL for 15 minutes at room temperature before cell isolation. Wash sample twice with isolation buffer, centrifuged at 300 g for 5 minutes. Filter aggregated suspensions through a 70 μm mesh nylon strainer and adjust cell density at 1×10^8 cells/mL.

➢ Cell Isolation

a) Prepare 100 μL of cell suspension (about 1×10^7 cells), add 30 μL Human $CD8^+$ T Cell Isolation Cocktail, mix fully and incubate for 5 minutes at room temperature.

   Note: Please ensure that the cells are in a single-cell suspension. Before sample dilution, filter the samples through a 70 μm cell sieve. The frozen PBMC sample need to be treated with DNase I and then filtered through a 70 μm cell sieve again before isolating.

b) Add isolation buffer to a final volume of 2 mL, centrifuge at 300 g for 5 minutes. Discard the supernatant, and then resuspend the cells with 100 μL isolation buffer.

   Note:

   - If the total volume of the cell suspension exceeds 1 mL, the volume of the added isolation buffer shall be no less than the total volume of the cell suspension.For example, if the total volume of the cell suspension is 1.5 mL, the volume of the isolation buffer added shall be ≥ 1.5 mL.
   - To maintain consistent cell density, the volume of cell isolation buffer for cell resuspension shall be identical to that of the input cell suspension. In the protocol example, if 100 μL of cell suspension is used as the starting input, cells should be resuspended with an equal volume of 100 μL cell isolation buffer.

c) Wash Beads Streptavidin 1.0-N: Place a clean flow cytometry tube or a centrifuge tube compatible with the magnetic rack into a tube rack. Pipette 1 mL of isolation buffer into the tube, vortex beads for 20 seconds, then add 24 μL of magnetic beads directly into the aforementioned 1 mL of isolation buffer. Mix by pipetting up and down 6-8 times. Place the flow cytometry tube or

centrifuge tube on a magnetic rack (provided by the user) and magnetically separate at room temperature for 5 min. At this point, the magnetic beads are attracted to the tube wall. Keep the tube on the magnetic rack, discard the supernatant, and then remove the tube from the magnetic rack.

Note：If the total volume of magnetic beads to be washed is greater than 1 mL, use a 1:1 volume ratio of isolation buffer to beads during the washing step.

d) Resuspend the magnetic beads using the cell suspension from step b): Aspirate the cell suspension and pipette the beads off the tube wall to the bottom of the tube (Note: avoid generating bubbles). Mix by pipetting up and down 6-8 times, then incubate at room temperature for 5 min.

Note:

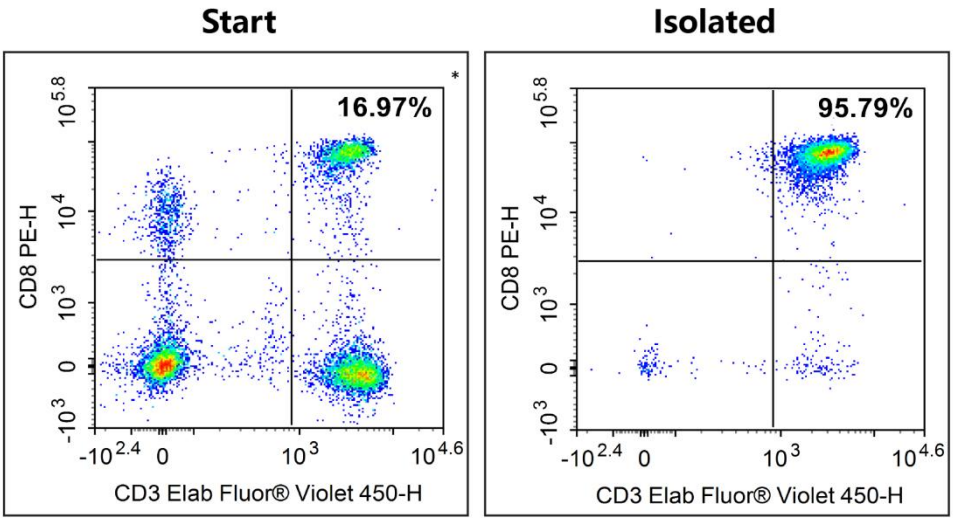
- If more than 1×10^7 cells are to be isolated, increase the amount of Human $CD8^+$ T Cell Isolation Cocktail and Human $CD8^+$ T Beads Streptavidin 1.0-N proportionally while ensuring the cell density remains 1×10^8 cells/mL. If fewer than 1×10^7 cells are to be isolated, resuspend the cells with 100 µL isolation buffer, add 30 µL Human $CD8^+$ T Cell Isolation Cocktail and 24 µL washed Human $CD8^+$ T Beads Streptavidin 1.0-N.
- The 5 mL flow tube is suitable for isolation of cell suspension ≤ 1 mL (1×108 cells). 10 mL or 15 mL centrifuge tube is suitable for isolation of cell suspension ≤ 4 mL (4×108 cells).

e) Add isolation buffer to a final volume of 2.5 mL (If the volume of the cell suspension for isolation is >1 mL, resuspend in an equal volume of isolation buffer), mix gently with a pipette by blowing up and down for 7-8 times until no particles of magnetic beads are visible. Put the tube on a 5 mL magnetic rack (self-provided) and stand for 5 minutes.

Note: Please mix the liquid thoroughly to avoid the magnetic beads clumping and affecting the isolation efficiency.

f) Transfer the cell suspension to a clean centrifuge tube, centrifuge at 300 g for 5 minutes. Discard the supernatant, resuspend the cells with buffer required for the subsequent experiments.

Typical data



The CD8⁺ T cells isolated from normal PBMC were stained with Elab Fluor® Violet 450 Anti-Human CD3 Antibody[OKT-3] (E-AB-F1001Q) and PE Anti-Human CD8a [HIT8a] (E-AB-F1271D) . The purities of the start and final isolated fractions were 16.97% and 95.79%, respectively.

Cautions

1. This kit is for research use only.
2. Please take safety precautions and follow the procedures of laboratory reagent operation.
3. Avoid freezing and thawing during the use and storage of the beads.
4. Sample differences, sample preparation and experimental operation have an important impact on the final isolated cell purity. The statistical results for cell purity in this product are based on normal human PBMC samples.
5. The quality of pre-isolated PBMC sample is critically impacts the separation efficiency of this product. It is recommended to test whether the percentage of $CD3^+CD8^+$ T cells is in the normal physiological range (5%-30%) after the preparation of PBMC sample. It is recommended to re-prepare the PBMC sample when percentage of target cell population is lower than it's normal distribution.
6. The cell clusters in the cell suspension will affect the purity of cell isolation. Therefore, cell suspension should be filtered with a 70 μm mesh nylon sieve before formal isolation.
7. Perform isolation immediately after preparing the cell suspension, as cell viability will decrease with longer storage time.
8. When adding the antibody cocktail and aspirating the magnetic beads for washing, pipette them directly to the bottom of the tube to avoid adhesion to the wall, which would result in loss of components.
9. In order to ensure the activity of the cells, the whole process of the experiment should be completed on ice as much as possible, except for the incubation at room temperature.
10. It is recommended to use low adsorption pipette tips and centrifuge tubes to avoid the loss of

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magnetic beads and antibodies due to adsorption.

11. The kit should be used in combination with a magnetic rack.

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