

(FOR RESEARCH USE ONLY. DO NOT USE IT IN CLINICAL DIAGNOSIS !)

**Catalog No: E-BC- K809-M**

**Specification: 48T(46 samples)/96T(94 samples)**

**Measuring instrument: Microplate reader (340 nm)**

**Detection range: 3.22-446.95 U/L**

## **Elabsience® Cell Glutathione Peroxidase (GPX) Activity Assay Kit**

This manual must be read attentively and completely before using this product.

If you have any problem, please contact our Technical Service Center for help:

Toll-free: 1-888-852-8623

Tell: 1-832-243-6086

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Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

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**Assay summary**



## **Intended use**

This kit can be used to measure glutathione peroxidase (GPX) activity in cell samples.

## **Detection principle**

Glutathione peroxidase (GPX) plays an important role in protecting cells from free radical damage caused by hydrogen peroxide decomposition. The lipid components of cells are prone to react with free radicals, leading to lipid peroxidation. GPX uses glutathione to reduce peroxides to alcohols, thereby preventing cell damage.

GPX catalyzes the substrate, and the product consumes the reducing. The reducing agent has a maximum absorbance at 340 nm. The activity of GPX can be calculated by measuring the change of absorbance value at 340 nm.

## Kit components & storage

Item	Component	Size 1(48 T)	Size 2(96 T)	Storage
Reagent 1	Buffer Solution	25 mL ×1 vial	50 mL × 1 vial	-20℃, 12 months
Reagent 2	Substrate	Powder × 1 vial	Powder × 2 vials	-20℃, 12 months, shading light
Reagent 3	Enzyme Reagent	0.25 mL ×1 vial	0.5 mL × 1 vial	-20℃, 12 months, shading light
Reagent 4	Oxidant	0.15 mL ×1 vial	0.3 mL × 1 vial	-20℃, 12 months, shading light
Reagent 5	Reductant	Powder × 1 vial	Powder × 2 vials	-20℃, 12 months, shading light
Reagent 6	Accelerant	4 mL ×1 vial	8 mL ×1 vial	-20℃, 12 months, shading light
Reagent 7	Stabilizer	0.5 mL ×1 vial	1 mL × 1 vial	-20℃, 12 months, shading light
Reagent 8	Cell Lysis Buffer	7 mL ×1 vial	14 mL × 1 vial	-20℃, 12 months
	UV Microplate	96 wells		No requirement
	Plate Sealer	2 pieces		
	Sample Layout Sheet	1 piece		

Note: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other. For a small volume of reagents, please centrifuge before use, so as not to obtain sufficient amount of reagents.

## Materials prepared by users

### Instruments:

Microplate reader (340 nm), Incubator

## Reagent preparation

① Equilibrate all reagents to room temperature before use.

② Preparation of substrate working solution:

Dissolve one vial of substrate with 250  $\mu\text{L}$  of double distilled water, mix well to dissolve. Store at  $-20^{\circ}\text{C}$  for 2 days protected from light.

③ Preparation of oxidant working solution:

Before testing, please prepare sufficient oxidant working solution according to the test wells. For example, prepare 100  $\mu\text{L}$  of oxidant working solution (mix well 95  $\mu\text{L}$  of buffer solution and 5  $\mu\text{L}$  of oxidant). Keep oxidant working solution on ice during use. The oxidant working solution should be prepared on spot and used up within 2 hours.

④ Preparation of reductant working solution:

Dissolve one vial of reductant with 500  $\mu\text{L}$  of double distilled water, mix well to dissolve. Store at  $-20^{\circ}\text{C}$  for 2 days protected from light.

⑤ Preparation of reaction working solution:

Before testing, please prepare sufficient reaction working solution according to the test wells. For example, prepare 1285  $\mu\text{L}$  of reaction working solution (mix well 825  $\mu\text{L}$  of buffer solution, 10  $\mu\text{L}$  of substrate working solution, 25  $\mu\text{L}$  of enzyme reagent, 25  $\mu\text{L}$  of reducing working solution and 400  $\mu\text{L}$  of accelerant). Keep reaction working solution on ice during use. The reaction working solution should be prepared on spot and used up within 2 hours.

## Sample preparation

### ① Sample preparation

#### Cell (adherent or suspension) samples:

- ① Harvest the number of cells needed for each assay (initial recommendation  $1 \times 10^6$  cells).
- ② Wash cells with PBS (0.01 M, pH 7.4).
- ③ Homogenize  $1 \times 10^6$  cells in 99  $\mu$ L cell lysis buffer and 1  $\mu$ L stabilizer. Crack on ice for 10 minutes, mix well every 5 minutes.
- ④ Centrifuge at  $10000 \times g$  for 10 min at  $4^\circ\text{C}$  to remove insoluble material. Collect supernatant and keep it on ice for detection.
- ⑤ Meanwhile, determine the protein concentration of supernatant (E-BC-K318-M).

### ② Dilution of sample

The recommended dilution factor for different samples is as follows (for reference only):

Sample type	Dilution factor
$1.9 \times 10^6$ HL-60 cells	1
$1.4 \times 10^6$ 293T cells	1
$2 \times 10^6$ CHO cells	1
$1.9 \times 10^6$ 4T1 cells	1
$2.03 \times 10^6$ Jurkat cells	1

Note: The diluent is cell lysis buffer. For the dilution of other sample types, please do pretest to confirm the dilution factor.

## The key points of the assay

If the  $A_2$  of sample well is less than 0.2, the sample must be diluted and test again.

## Operating steps

- ① Sample well: Add 20  $\mu\text{L}$  of sample into sample well.  
Blank well: Add 20  $\mu\text{L}$  of cell lysis buffer into blank well.
- ② Add 180  $\mu\text{L}$  of reaction working solution into each well.
- ③ Add 40  $\mu\text{L}$  of oxidant working solution into each well.
- ④ Mix fully with microplate reader for 5 s. After adding oxidant working solution, measure the OD values of each well at 340 nm with microplate reader immediately recorded as  $A_1$ . Incubate at room temperature for 10 min, measure the OD values of each well at 340 nm recorded as  $A_2$ . As the reaction rate is fast, If  $A_1$  is not determined in time, it may lead to low or no value in the sample.



## Calculation

### Cell sample:

**Definition:** The amount of GPX in 1 g cell protein per minute that catalyze the substrate to produce 1  $\mu\text{mol}$  substance at 25°C is defined as 1 unit.

$$\text{GPX activity (U/gprot)} = (\Delta A_{\text{sample}} - \Delta A_{\text{blank}}) \div (\epsilon \times d) \times (V_{\text{total}} \div V_{\text{sample}}) \div T \times f \div C_{\text{pr}}$$

### [Note]

$\Delta A_{\text{sample}}$ : The change OD value of sample well ( $\Delta A = A_1 - A_2$ )

$\Delta A_{\text{blank}}$ : The change OD value of blank well ( $\Delta A = A_1 - A_2$ )

$\epsilon$ : the molar extinction coefficient of product at 340 nm,  $6.22 \times 10^3$  L/ $\mu\text{mol}/\text{cm}$

d: Optical path, 0.6 cm.

$V_{\text{total}}$ : the total volume of reaction, 0.24 mL.

$V_{\text{sample}}$ : the volume of sample, 0.02 mL.

T: The time of reaction, 10 min.

$C_{\text{pr}}$ : The concentration of protein in sample, gprot/L.

f: Dilution factor of sample before test.

## Appendix I Performance Characteristics

### 1. Parameter:

#### Intra-assay Precision

Three HL-60 cell samples were assayed in replicates of 20 to determine precision within an assay (CV = Coefficient of Variation).

Parameters	Sample 1	Sample 2	Sample 3
Mean (U/gprot)	98.53	116.90	129.05
%CV	3.10	1.20	1.50

#### Inter-assay Precision

Three HL-60 cell samples were assayed 20 times in duplicate by three operators to determine precision between assays.

Parameters	Sample 1	Sample 2	Sample 3
Mean (U/gprot)	79.97	66.10	58.03
%CV	0.40	3.10	3.40

#### Recovery

Take three samples of high concentration, middle concentration and low concentration to test the samples of each concentration for 6 times parallelly to get the average recovery rate of 100%.

	Sample 1	Sample 2	Sample 3
Expected Conc. (U/gprot)	102.29	116.90	131.51
Observed Conc. (U/gprot)	98.53	116.90	129.05
Recovery rate (%)	96.33	100.00	98.13

#### Sensitivity

The analytical sensitivity of the assay is 3.22 U/L. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 20 times, and calculating the corresponding concentration.

## Appendix Π Example Analysis

### Example analysis :

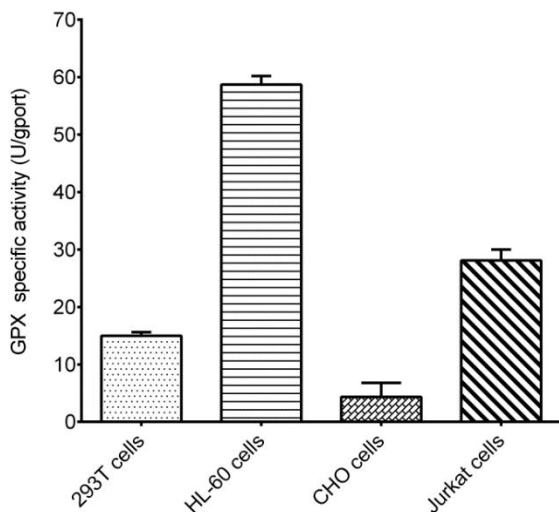
For HL-60 cell, take 20  $\mu\text{L}$  of supernatant and carry the assay according to the operation table. The results are as follows:

The  $A_1$  of the sample is 1.153, the  $A_2$  of the sample is 0.851,  $\Delta A_{\text{sample}} = A_1 - A_2 = 1.153 - 0.851 = 0.302$ . The  $A_1$  of the blank is 1.163, the  $A_2$  of the blank is 1.157,  $\Delta A_{\text{blank}} = A_1 - A_2 = 1.163 - 1.157 = 0.006$ . The concentration of protein in sample is 1.698 gprot/L and the calculation result is:

GPX activity (U/gprot)

$$= (0.302 - 0.006) \div (6.22 \times 10^{-3} \times 0.6) \times (0.24 \div 0.02) \div 10 \div 1.698 = 56.05 \text{ U/gprot}$$

Detect 293T cell (the concentration of protein is 1.235 gprot/L), HL-60 cell (the concentration of protein is 1.698 gprot/L), CHO cell (the concentration of protein is 1.850 gprot/L) and Jurkat cell (the concentration of protein is 0.68 gprot/L) according to the protocol, the result is as follows:



## Statement

1. This assay kit is for Research Use Only. We will not response for any arising problems or legal responsibilities causing by using the kit for clinical diagnosis or other purpose.
2. Please read the instructions carefully and adjust the instruments before the experiments. Please follow the instructions strictly during the experiments.
3. Protection methods must be taken by wearing lab coat and latex gloves.
4. If the concentration of substance is not within the detection range exactly, an extra dilution or concentration should be taken for the sample.
5. It is recommended to take a pre-test if your sample is not listed in the instruction book.
6. The experimental results are closely related to the situation of reagents, operations, environment and so on. Elabscience will guarantee the quality of the kits only, and NOT be responsible for the sample consumption caused by using the assay kits. It is better to calculate the possible usage of sample and reserve sufficient samples before use.