LIN (Lincomycin) ELISA Kit

Catalog No: E-FS-E061

96T

Version Number:	V1.2
Replace version:	V1.1
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This manual must be read attentively and completely before using this product.

If you have any problems, please contact our Technical Service Center for help.

Toll-free: 1-888-852-8623 Tel: 1-832-243-6086 Fax: 1-832-243-6017
Email: techsupport@elabscience.com
Website: www.elabscience.com

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

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Test principle

This kit uses Competitive-ELISA as the method for the quantitative detection. It can detect lincomycin (LIN) in samples, such as honey, urine, tissue, etc. This kit is composed of ELISA Microtiter plate, HRP conjugate, antibody working solution, standard and other supplementary reagents. The microtiter plate provided in this kit has been pre-coated with coupled antigen. During the reaction, LIN in the samples or standard competes with coupled antigen on the solid phase supporter for sites of anti-LIN antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each microtiter plate well, and substrate reagent is added for color development. There is a negative correlation between the OD value of samples and the concentration of LIN. The concentration of LIN in the samples can be calculated by comparing the OD of the samples to the standard curve.

Technical indicators

Reaction mode(Incubation time and temperature)**:** 25℃; 30 min, 15 min.

Detection limit: Urine, honey---1.5 ppb; tissue---3 ppb.

Cross-reactivity: Lincomycin---100%

Sample recovery rate: Urine ---90% ±10%; Muscle --- 80% ±10%; Honey --- 80% ±15%.

Kit components

Item	Specifications
ELISA Microtiter plate	96 wells
Standard Liquid	1 mL each (ppb=ng/mL=ng/g) (0 ppb, 0.05 ppb, 0.15 ppb, 0.45 ppb, 1.35 ppb, 4.05 ppb)
HRP Conjugate	5.5 mL
Antibody Working Solution	5.5 mL
Substrate Reagent A	6 mL
Substrate Reagent B	6 mL
Stop Solution	6 mL
20×Concentrated Wash Buffer	40 mL
2×Reconstitution Buffer	50 mL
Plate Sealer	1 piece
Sealed Bag	1 piece
Manual	1 copy

Note: All reagent bottle caps must be tightened to prevent evaporation and microbial pollution.

Other materials required but not supplied

Instrument: Microplate reader, Printer, Homogenizer, Vortex mixer, Centrifuge, Graduated pipette, Balance (sensibility 0.01 g).

Micropipette: Single channel (20-200 μL, 100-1000 μL), Multichannel (30-300 μL).

Reagents: Methanol, HCl.

Notes

1. The overall OD value will be lower when reagents have not been brought to room temperature before use or room temperature is below 25℃.
2. If the wells turn dry during the washing procedure, it will lead to bad linear standard curve and poor repeatability. Operate the next step immediately after wash.
3. Mix thoroughly and wash the plate completely. The consistency of wash procedure can strongly affect the reproducibility of this ELISA kit.
4. FOR RESEARCH USE ONLY. ELISA Microtiter plate should be covered by plate sealer. Avoid th e kit to strong light.
5. **Each reagent is optimized for use in the E-FS-E061. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other E-FS-E061 with different lot numbers.**
6. Substrate Reagent should be abandoned if it turns blue color. When OD value of standard (concentration: 0) < 0.5 unit (A450nm < 0.5), it indicates the reagent be deteriorated.
7. Stop solution is caustic, avoid contact with skin and eyes.
8. As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.
9. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
10. **For mentioned sample fast and efficient extraction methods are included in the kit description. Please consult technical support for the applicability if other sample need to be tested.**
11. The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS, etc. can be used for quantitative confirmation.

Storage and expiry date

Store the kit at 2-8℃. Do not freeze any test kit components.

Return any unused microwells to their original foil bag and reseal them together with the desiccant provided and further store at 2-8℃. After opening, the kit is stable for up to 1 month.

Expiry date: expiration date is on the packing box.

Experimental preparation

Restore all reagents and samples to room temperature before use.

Open the microplate reader in advance, preheat the instrument, and set the testing parameters.

1. **Sample pretreatment Notice:**

   Experimental apparatus should be clean, and the pipette should be disposable to avoid cross-contamination during the experiment.

2. **Solution preparation**

   Please prepare solution according to the number of samples. Don't use up all components in the kit at once!

   Solution 1: **Extraction Solution**

   Dissolve 0.75 mL of **HCl** and 100 mL of **Methanol** with 600 mL of deionized water.

   Solution 2: **Reconstitution Buffer**

   Dilute the **2×Reconstitution Buffer** with deionized water. (2×Reconstitution Buffer (V): Deionized water (V)=1:1) .The Reconstitution buffer can be store at 4℃ for a month.

   Solution 3: **Wash Buffer**

   Dilute the **20×Concentrated Wash Buffer** with deionized water. (20×Concentrated Wash Buffer (V): Deionized water (V) =1:19).

3. **Sample pretreatment**

3.1 Pretreatment of urine (swine) sample:

(1) Take 100 µL of clear urine to centrifuge tube (If the urine sample is turbid, it should be filtered or 4000 rpm for 5 minutes until the urine sample is clear, take the supernatant for testing).

(2) And add 900 µL of **Reconstitution Buffer** (Solution 2) into centrifuge tube, mix fully.

(3) Take 50 µL for analysis.

Note: Sample dilution factor: 10, detection limit: 1.5 ppb.

3.2 Pretreatment of muscle (livestock) sample:

(1) Remove fat from sample, homogenize the sample with homogenizer.

(2) Weigh 1±0.05 g of homogenate sample into a centrifuge tube, add 5 mL of **Extraction Solution** (Solution 1) and vortex for 3 min. Centrifuge at 4000 r/min for 10min at room temperature.

(3) Take 0.2 mL of the supernatant to another centrifuge tube, add 0.6 mL of **Reconstitution Buffer** (Solution 2), and mix fully for 30 s.

(4) Take 50 µL for analysis.

Note: Sample dilution factor: 20, detection limit: 3 ppb.

3.3 Pretreatment of honey sample:

(1) Weigh 1±0.05 g of honey sample into centrifuge tube, dissolved with 5 mL of deionized water and vortex for 2 min. Centrifuge at 4000 r/min for 5 min at room temperature.

(2) Take 1 mL of supernatant with 1 mL of **Reconstitution Buffer** (Solution 2) to another centrifuge tube, mix fully.

(3) Take 50 µL for analysis.

Note: Sample dilution factor: 10, detection limit: 1.5 ppb.

Assay procedure

Restore all reagents and samples to room temperature before use. All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming. The unused ELISA Microtiter plate should be sealed as soon as possible and stored at 2-8℃.

1. **Number:** number the sample and standard in order (multiple well), and keep a record of standard wells and sample wells. **Standard and Samples need test in duplicate.**
2. **Add Sample:** add 50 µL of **Standard** or **Sample** per well, add 50 µL of **HRP conjugate** to each well, then add 50 µL **Antibody Working Solution**, cover the plate with plate sealer. Oscillate for 5 s gently to mix thoroughly. Incubate at 25℃ for 30 min with shading light.
3. **Wash:** uncover the sealer carefully, remove the liquid in each well. Immediately add 350 µL of **Wash Buffer** (Solution 3) to each well and wash. Repeat wash procedure for 5 times, 30 s intervals/time. Invert the plate and pat it against thick clean absorbent paper (If bubbles exist in the wells, clean tips can be used to prick them).
4. **Color Development:** add 50 µL of **Substrate Reagent A** to each well, and then add 50 µL of **Substrate Reagent B**. Gently oscillate for 5 s to mix thoroughly. Incubate at 25℃ for 15 min with shading light (The reaction time can be extended according to the actual color change).
5. **Stop Reaction:** add 50 µL of **Stop Solution** to each well, oscillate gently to mix thoroughly.
6. **OD Measurement:** determine the optical density (OD value) of each well at 450 nm (reference wavelength 630 nm) with a microplate reader. This step should be finished in 10 min after stop reaction.

Result analysis

1. **Absorbance (%)=A/A_0×100%**

   A: Average absorbance of standard or sample

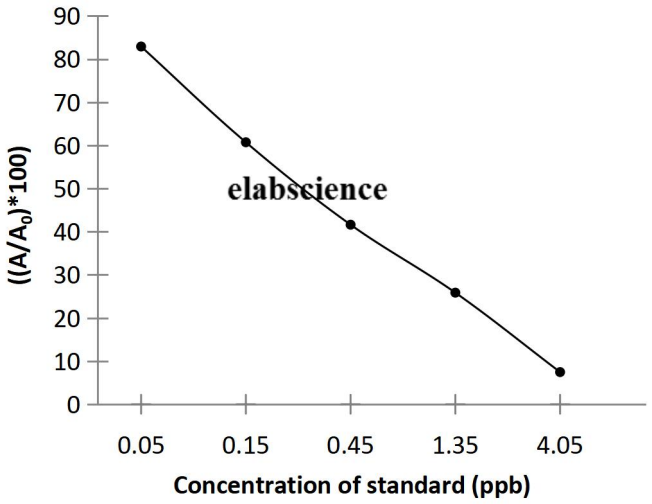
   A_0: Average absorbance of 0 ppb Standard

2. **Drawing and calculation of standard curve**

   Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add average absorbance value of sample to standard curve to get corresponding concentration. **If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.**

   For this kit, it is more convenient to use professional analysis form for accurate and fast analysis on a large number of samples.

Lincomycin (E-FS-E061) **Standard Curve**



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