（本试剂盒仅供体外研究使用，不用于临床诊断!）

产品货号：GEOSH0057

产品规格：96T/48T/24T/96T*5

Elabscience®QuicKey Pro-人转铁蛋白受体 1（TFR1）酶联免疫吸附测定试剂盒使用说明书

QuicKey Pro Human TFR1 (Transferrin Receptor 1) ELISA Kit

使用前请仔细阅读说明书。如果有任何问题，请通过以下方式联系我们：

电话：400-999-2100

邮箱：techsupport@elabscience.cn

网址：www.elabscience.cn

具体保质期请见试剂盒外包装标签。请在保质期内使用试剂盒。

联系时请提供产品批号(见试剂盒标签)，以便我们更高效地为您服务。

QuicKey Pro™系列

与传统ELISA试剂盒相比，QuicKey Pro™系列在实验时间节省至少1-2小时的同时获得更加灵敏和精确的实验结果。Elabscience® 自主开发的新技术，旨在帮助客户以更为高效的方式进行科学研究。

用途

该试剂盒用于体外定量检测人血清和血浆中TFR1浓度，其余样本类型检测请咨询技术支持。

基本性能

性能	
灵敏度	0.23 ng/mL
检测范围	0.39-25 ng/mL
特异性	可检测样本中的人 TFR1，且与其它类似物无明显交叉反应
重复性	板内，板间变异系数均<10%

检测原理

本试剂盒采用双抗体夹心 ELISA 法。用抗人 TFR1 抗体包被于酶标板上，实验时加入样品（或标准品）和辣根过氧化物酶标记的抗人 TFR1 检测抗体，样本(或标准品中)的人 TFR1 会与包被抗体及辣根过氧化物酶标记的抗人 TFR1 抗体结合，形成双抗体夹心的免疫复合物，游离的成分被洗去。加入显色底物(TMB)，TMB 在辣根过氧化物酶的催化下呈现蓝色，加终止液后变成黄色。用酶标仪在 450 nm 波长处测 OD 值，TFR1 浓度与 OD_{450} 值之间呈正比，通过绘制标准曲线计算出样品中 TFR1 的浓度。

试剂盒组成及保存

未拆封的试剂盒可在 2-8℃保存 **6 个月**；试剂盒组分开封后请按照下表中的条件分别保存。

中文名称 Micro ELISA Plate	规格	开封后保存条件
ELISA 酶标板 Micro ELISA Plate	96T：8 孔×12 条 48T：8 孔×6 条 24T：8 孔×3 条 96T*5：5 块 96T 酶标板	2-8℃，可存放 1 个月
冻干标准品 Reference Standard	96T：2 支 48T/24T：1 支 96T*5：10 支	2-8℃，复溶后当天使用
浓缩 HRP 酶结合物（100×） Concentrated HRP Conjugate（100×）	96T：1 支 60μL 48T/24T：1 支 30μL 96T*5：5 支 60μL	2-8℃(避光)
标准品&样品稀释液 Reference Standard & Sample Diluent	96T/48T/24T：2 瓶 20mL 96T*5：10 瓶 20mL	2-8℃
酶结合物稀释液 HRP Conjugate Diluent	96T/48T/24T：1 瓶 14mL 96T*5：5 瓶 14mL	
浓缩洗涤液 (25×) Concentrated Wash Buffer (25×)	96T/48T/24T：1 瓶 30mL 96T*5：5 瓶 30mL	
底物溶液(TMB) Substrate Reagent	96T/48T/24T：1 瓶 10mL 96T*5：5 瓶 10mL	2-8℃(避光)
反应终止液 Stop Solution	96T/48T/24T：1 瓶 10mL 96T*5：5 瓶 10mL	2-8℃
封板覆膜 Plate Sealer	96T/48T/24T：5 张 96T*5：25 张	
产品说明书 Manual	1 份	
质检报告 Certificate of Analysis	1 份	

说明：所有试剂瓶盖须旋紧以防止蒸发和微生物的污染。

试剂体积以实际发货版说明书为准。相关试剂在分装时会比标签上标明的体积稍多一些，请在使用时量取而非直接倒出。

试验所需自备物品

1. 酶标仪(450 nm波长滤光片)
2. 高精度移液器，EP管及一次性吸头：0.5-10μL, 2-20μL, 20-200μL, 200-1000μL
3. 37℃恒温箱
4. 双蒸水或去离子水
5. 吸水纸
6. 加样槽

样品收集方法

1. 血清：全血样品于室温放置2小时或4℃过夜后于1000×g离心20分钟，取上清即可检测，收集血液的试管应为一次性的无内毒素试管。
2. 血浆：抗凝剂推荐使用EDTA-Na2，样品采集后30分钟内于1000×g离心15分钟，取上清即可检测。避免使用溶血，高血脂样品。

注意事项

■ 试剂盒注意事项

1) 本试剂盒仅供体外研究使用，不用于临床诊断。
2) 试验中请穿着实验服并戴乳胶手套做好防护工作。特别是检测血液或者其他体液样品时，请按国家生物试验室安全防护条例执行。
3) 刚开启的酶标板孔中可能会有少许水样物质，此为正常现象，不会对实验结果造成任何影响。暂时不用的板条应拆卸后放入备用铝箔袋，按照上述表格中保存条件存放。
4) 请勿重复使用已稀释过的标准品、HRP酶结合物工作液。未用完的浓缩HRP酶结合物(100×)、酶标板及其他原液按照上述表格中保存条件存放。
5) 检测使用的酶标仪需要安装能检测450±2 nm波长的滤光片，光密度范围在0-3.5之间。建议使用时提前15分钟预热。
6) 请勿使用其他批号或其他来源的试剂混合或替代本试剂盒中的试剂。
7) 试验中所用的EP管和吸头均为一次性使用，严禁混用。
8) 请勿使用过期的试剂。

■ 样品注意事项

1) 收集血液的试管应为一次性无内毒素试管。避免使用溶血，高血脂样品。
2) 样品收集后若在1周内进行检测可保存于2-8℃，若不能及时检测，请按一次使用量分装，冻存于-20℃(1个月内检测)，或-80℃(3个月内检测)，避免反复冻融。在检测前，冷冻过的样本应缓慢地融化并离心除去冻融过程产生的沉淀物。室温混匀后使用。
3) **试剂盒检测范围不等同于样本中待测物的浓度范围**。如果样品中待测物浓度过高或过低，请对样本做适当的稀释或浓缩。
4) 若所检样本不在说明书所列样本之中，建议做预实验验证其检测有效性。若使用化学裂解液制备组织匀浆或细胞提取液，由于引入某些化学物质会导致ELISA测值出现偏差。
5) 某些重组蛋白可能与试剂盒中捕获或检测抗体不匹配而出现不能检测的情况。

样本稀释方案

人血清、血浆建议稀释 200-1000 倍检测。

由于存在个体差异，请提前预估样本的浓度范围，并通过预实验确定待检样本的稀释倍数。

如果您的检测样本需要稀释，通用稀释方案参考如下：

稀释100倍：一步稀释。取5 μL样本到495 μL标准品&样本稀释液内，做100倍稀释；

稀释 1000 倍：两步稀释。取 5 μL 样本到 95 μL 标准品&样本稀释液内，做 20 倍稀释，再取 5 μL 20 倍稀释样本到 245 μL 标准品&样本稀释液内，做 50 倍稀释，总共稀释 1000 倍；

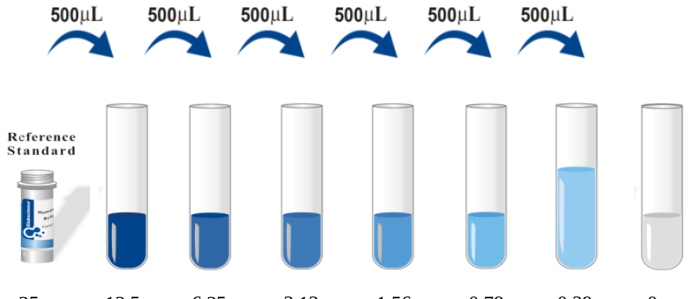
稀释 100000 倍：三步稀释。取 5 μL 样本到 195 μL 标准品&样本稀释液内，做 40 倍稀释，再取 5 μL 40 倍稀释样本到 245 μL 标准品&样本稀释液内，做 50 倍稀释，最后取 5 μL 2000 倍稀释样本到 245 μL 标准品&样本稀释液内，做 50 倍稀释，总共稀释 100000 倍；

每步稀释时取液量不少于 **3 μL**，稀释倍数不超过 **100 倍**。每步稀释都需混合均匀，避免起泡。

检测前准备工作

1. 提前20分钟从冰箱中取出试剂盒，平衡至室温(18-25℃)。如果试剂盒需分多次使用，请仅取出本次实验所需的酶标板条和试剂，剩余板条和试剂需按照指定条件保存。
2. **洗涤液：**将**浓缩洗涤液**用双蒸水稀释(1:24)。提示：从冰箱中取出的浓缩洗涤液可能有结晶，属于正常现象，可用40℃水浴微加热使结晶完全溶解后再配制洗涤液。当日使用。
3. **标准品工作液：**将**标准品**于10000×g离心1分钟，加入**标准品&样品稀释液** 1 mL至冻干标准品中，旋紧管盖，静置10分钟，上下颠倒数次，待其充分溶解后，轻轻混匀，避免起泡，配成25 ng/mL的标准品工作液(或加入1 mL **标准品&样品稀释液**后，静置1-2分钟，用低速涡旋仪充分混匀。可通过低速离心去除涡旋过程中产生的气泡)。然后根据需要进行倍比稀释。建议配制成以下浓度：25、12.5、6.25、3.13、1.56、0.78、0.39、0 ng/mL。
   倍比稀释方法：取7支EP管，每管中加入500 μL**标准品&样品稀释液**，从25 ng/mL的标准品工作液中吸取500 μL到第一支EP管中混匀配成12.5 ng/mL的标准品工作液，按此步骤往后依次吸取混匀。如下页图示。提示：最后一管直接作为空白孔，不需要再从倒数第二管中吸取液体。倍比稀释的标准品工作液需要现配现用。
4. **HRP酶结合物工作液：**HRP酶结合物为HRP酶结合抗体。实验前计算当次实验所需用量(以50 μL/孔计算)，实际配制时应多配制100-200 μL。使用前15分钟，将**浓缩HRP酶结合物**于800×g离心1分钟，以**酶结合物稀释液**将100×**浓缩HRP酶结合物**稀释成1×工作浓度(例如：10 μL浓缩液+990 μL稀释

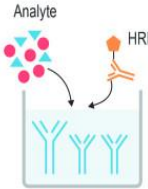
液)。现配现用。



操作步骤

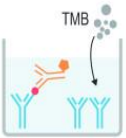
1. 分别设定**标准孔**、**空白孔**和**样本孔**。标准孔加入 50 μL 倍比稀释的标准品，空白孔加入 50 μL 标准品&样本稀释液，其余孔加入 50 μL 待测样本(建议所有的待检样本和标准品在检测中设立复孔；建议通过预实验或咨询技术支持确定待检样本的稀释倍数)。随后每孔立即加入 **HRP 酶结合物工作液** 50 μL，酶标板加上覆膜，37℃温育 60 分钟。提示：加样时将样品加于酶标板底部，尽量不触及孔壁，轻轻晃动混匀，避免产生气泡。整个加样时间宜控制在 10 分钟内。
2. 甩尽孔内液体，在洁净的吸水纸上拍干。每孔加**洗涤液** 350 μL，浸泡 1 分钟，吸去或甩掉酶标板内的液体，拍干。重复此洗板步骤 5 次。提示：此处与其他洗板步骤都可使用洗板机(参考北京拓普 DEM-3 型洗板机参数设置：2 点吸，每孔加入洗涤液 350 μL，振板 5 秒，吸液 0.5 秒)。洗板完成后请立即进行下步操作，不要让微孔板干燥。
3. 每孔加**底物溶液(TMB)** 90 μL，酶标板加上覆膜，37℃避光孵育 15 分钟左右。提示：根据实际显色情况酌情缩短或延长，但不可超过 30 分钟。当标准孔出现明显梯度时(前 4 个显色孔出现明显蓝色梯度)，即可终止。提前 15 分钟打开酶标仪预热。
4. 每孔加**终止液** 50 μL，终止反应。提示：终止液的加入顺序应尽量与底物溶液的加入顺序相同。
5. 立即用酶标仪在 450 nm 波长测量各孔的光密度(OD 值)。

操作一览表



1. 对应板孔中加入50μL标准品工作液或样本后，立即每孔加入50μL HRP酶标抗体工作液，37℃孵育60分钟

2. 弃掉板内液体，洗板5次



3. 每孔加入90μL底物溶液，37℃孵育15分钟左右

4. 每孔加入50μL终止液

5. 立即在450nm波长下读数，处理数据

结果判断

1. 计算标准品和样本复孔的平均OD值并减去空白孔的OD值作为校正值。以浓度为横坐标，OD值为纵坐标，在双对数坐标轴上拟合四参数逻辑函数的标准曲线。
2. 若样品OD值高于标准曲线上限，应适当稀释后重测并在计算样本浓度时乘以相应的稀释倍数。

技术资源

微信扫描左下二维码，获得更详细的ELISA实验指南和常规问题分析。

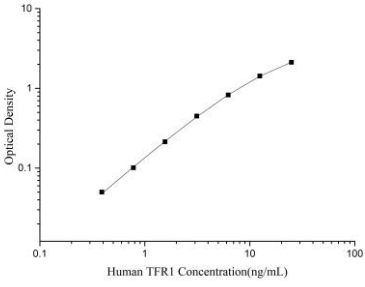
如有任何技术问题，请与我司技术支持联系(建议及时对显色结果拍照，保留实验数据、所用板条及未使用的试剂)。

ELISA实验指南

技术支持微信

典型数据

以下数据和曲线仅供参考，实验者需根据自己的实验建立标准曲线。

ng/mL	OD	校正 OD	标曲
25	2.196	2.120	
12.5	1.498	1.422	
6.25	0.900	0.824	
3.13	0.523	0.447	
1.56	0.289	0.213	
0.78	0.177	0.101	
0.39	0.126	0.050	
0	0.076	-	

样本测值

样本类型	样本中人TFR1浓度范围(ng/mL)
血清(n=10)	400-2000
血浆(EDTA)(n=10)	500-3000

注意：以上样本检测数据均来源于非生理期或妊娠期的正常个体。

性能

■ 精密度

板内精密度：低，中，高浓度样本分别在1块板子上检测20次。
板间精密度：低，中，高浓度样本分别在3块板子上检测20次。

	板内精密度			板间精密度		
样本	1	2	3	1	2	3
数量	20	20	20	20	20	20
平均值(ng/mL)	1.26	2.27	8.84	1.25	2.42	8.91
标准差	0.07	0.13	0.38	0.09	0.15	0.54
变异系数 (%)	5.61	5.71	4.35	7.12	6.1	6.01

■ 回收率

分别往不同样本中添加已知浓度的人TFR1，做回收实验，得出回收率范围和平均回收率。

样本类型	回收率范围 (%)	平均回收率 (%)
血清(n=8)	91-104	96
血浆(EDTA)(n=8)	93-108	101

■ 线性

将添加有人TFR1的样本分别稀释2倍，4倍，8倍，16倍做回收实验，得出回收率范围及平均回收率。

		血清 (n=5)	血浆(EDTA) (n=5)
1:2	回收率范围(%)	95-111	95-106
	平均回收率(%)	102	100
1:4	回收率范围(%)	96-113	89-103
	平均回收率(%)	104	95
1:8	回收率范围(%)	95-107	95-109
	平均回收率(%)	101	102
1:16	回收率范围(%)	95-111	89-102
	平均回收率(%)	101	96

声明

1. 限于现有条件及科学技术水平，尚不能对所有原料进行全面的鉴定分析，本产品可能存在一定的质量技术风险。
2. 本试剂盒在研发过程中去除/降低了生物学样本中的一些内源性干扰因素，并非所有可能影响的因素均已去除。
3. 最终的实验结果与试剂的有效性、实验者的相关操作以及当时的实验环境等因素密切相关，本公司只对试剂盒本身负责，不对因使用试剂盒所造成的样本消耗负责，请使用者使用前充分考虑到样本可能的使用量，预留充足的样本。
4. 为了达到好的实验结果，请只使用本公司试剂盒内提供的试剂，不要混用其他制造商的产品，严格按照说明书操作。
5. 由于操作过程中试剂制备以及酶标仪参数设置不正确，可能导致结果异常，实验前请仔细阅读说明书并调整好仪器。
6. 即使是相同人员操作也可能在两次独立实验中得到不同的结果，为保证结果的重现性，需要控制实验过程中每一步的操作。
7. 试剂盒发货前会经过严格的质检，然而，因为运输条件、实验设备差异等等因素影响，用户检测结果可能跟出厂数据不一致。不同批次间试剂盒间的差异也可能来自上述原因。
8. 本试剂盒未与其他厂家同类试剂盒或不同方法检测同一目的物的产品进行对比，所以不排除检测结果不一致的情况。
9. 试剂盒仅供研究使用，如将其用于临床诊断或任何其他用途，我公司将不对因此产生的问题负责，亦不承担任何法律责任。

QuicKey Pro Human TFR1 (Transferrin Receptor 1) ELISA Kit

Catalog No: GEOSH0057
Size: 96T/48T/24T/96T*5

QuicKey Pro™ Series

Get more sensitive and precise results with saving at least 1-2h comparing to traditional ELISA Kits. The new developed technology in house will help to accelerate your science research in a more efficient way.

Intended use

This ELISA kit applies to the in vitro quantitative determination of TFR1 concentrations in Human serum and plasma samples. Please contact tech-support for other sample type detection.

Character

Item	
Sensitivity	0.23 ng/mL
Detection Range	0.39-25 ng/mL
Specificity	This kit recognizes Human TFR1 in samples. No significant cross-reactivity or interference between Human TFR1 and analogues was observed
Repeatability	Coefficient of variation is < 10%

Test principle

This ELISA kit uses the Sandwich-ELISA principle. The micro ELISA plate provided in this kit has been pre-coated with an antibody specific to Human TFR1. Samples (or Standards) and Horseradish Peroxidase (HRP) linked antibody specific for Human TFR1 are added to the micro ELISA plate wells. Human TFR1 in samples (or standards) combines with the coated antibody and HRP linked detection antibody special to Human TFR1. Excess conjugate and unbound sample or standard are washed from the plate. The substrate solution is added to each well. The enzyme-substrate reaction is terminated by the addition of stop solution and the color turns yellow. The optical density (OD) is measured spectrophotometrically at a wavelength of 450±2 nm. The OD value is proportional to the concentration of Human TFR1. The concentration of Human TFR1 in the samples is then determined by comparing the OD of the samples to the standard curve.

Kit components & Storage

An unopened kit can be stored at 2-8℃ for 6 months. After test, the unused wells and reagents should be stored according to the table.

Item	Specifications	Storage After Preparation
Micro ELISA Plate (Dismountable)	96T: 8 wells ×12 strips 48T: 8 wells ×6 strips 24T: 8 wells ×3 strips 96T*5: 5 plates, 96T	2-8℃, 1 month
Reference Standard	96T: 2 vials 48T/24T: 1 vial 96T*5: 10 vials	2-8℃, use the reconstituted standard within 24h
Concentrated HRP Conjugate (100×)	96T: 1 vial, 60 μL 48T/24T: 1 vial, 30 μL 96T*5: 5 vials, 60 μL	2-8℃(Protect from light)
Reference Standard & Sample Diluent	96T/48T/24T: 2 vials, 20 mL 96T*5: 10 vials, 20 mL	2-8℃
HRP Conjugate Diluent	96T/48T/24T: 1 vial, 14 mL 96T*5: 5 vials, 14 mL	
Concentrated Wash Buffer(25×)	96T/48T/24T: 1 vial, 30 mL 96T*5: 5 vials, 30 mL	
Substrate Reagent	96T/48T/24T: 1 vial, 10 mL 96T*5: 5 vials, 10 mL	2-8℃(Protect from light)
Stop Solution	96T/48T/24T: 1 vial, 10 mL 96T*5: 5 vials, 10 mL	2-8℃
Plate Sealer	96T/48T/24T: 5 pieces 96T*5: 25 pieces	
Product Description	1 copy	
Certificate of Analysis	1 copy	

Note: All reagent bottle caps must be tightened to prevent evaporation and microbial pollution. The volume of reagents in partial shipments is a little more than the volume marked on the label, please use accurate measuring equipment instead of directly pouring into the vial(s).

Other supplies required

Microplate reader with 450 nm wavelength filter
High-precision transfer pipette, EP tubes and disposable pipette tips
Incubator capable of maintaining 37℃
Deionized or distilled water
Absorbent paper
Loading slot

Sample collection

Serum: Allow samples to clot for 2 hours at room temperature or overnight at 4℃ before centrifugation for 20 min at 1000×g at 2~8℃. Collect the supernatant to carry out the assay. Blood collection tubes should be disposable and be non-endotoxin.
Plasma: Collect plasma using EDTA-Na2 as anticoagulant. Centrifuge samples for 15 min at 1000×g at 2~8℃ within 30 min of collection. Collect the supernatant to carry out the assay. Hemolysed samples are not suitable for ELISA assay!

Note

■ Note for kit

1) For research use only. Not for use in diagnostic procedures.
2) Please wear lab coats, eye protection and latex gloves for protection. Please perform the experiment following the national security protocols of biological laboratories, especially when detecting blood samples or other bodily fluids.
3) A freshly opened ELISA plate may appear a water-like substance, which is normal and will not have any impact on the experimental results. Return the unused wells to the foil pouch and store according to the conditions suggested in the above table.
4) Do not reuse the reconstituted standard, HRP Conjugate working solution. The unspent undiluted HRP Conjugate (100×) and other stock solutions should be stored according to the storage conditions in the above table.
5) The microplate reader should be able to be installed with a filter that can detect the wave length at 450±2 nm. The optical density should be within 0-3.5. Follow the Instructions of the Microplate Reader for set-up and preheat it for 15 min before OD measurement.
6) Do not mix or substitute reagents with those from other lots or sources.
7) Change pipette tips in between adding of each standard level, between sample adding and between reagent adding. Also, use separate reservoirs for each reagent.
8) The kit should not be used beyond the expiration date on the kit label.

■ Note for sample

1) Tubes for blood collection should be disposable and be non-endotoxin. Samples with high hemolysis or much lipid are not suitable for ELISA assay.
2) Samples should be assayed within 7 days when stored at 2-8℃, otherwise samples must be divided up and stored at -20℃ (≤1 month) or -80℃ (≤3 months). Avoid repeated freeze-thaw cycles. Prior to assay, the frozen samples should be slowly thawed and centrifuged to remove precipitates.
3) Please predict the concentration before assaying. If the sample concentration is not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments.
4) If the sample type is not included in the manual, a preliminary experiment is suggested to verify the validity. If a lysis buffer is used to prepare tissue homogenates or cell lysates, there is a possibility of causing a deviation due to the introduced chemical substance.
5) Some recombinant protein may not be detected due to a mismatching with the coated antibody or detection antibody.

Dilution method

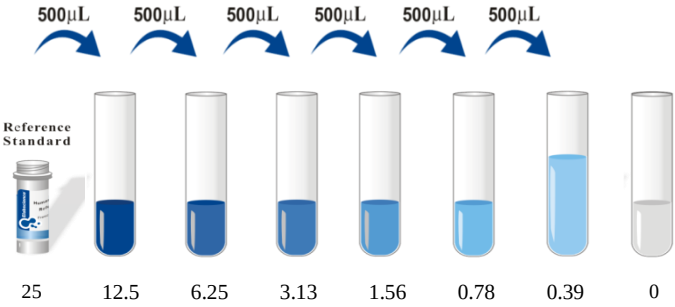
It is recommended to dilute normal serum/plasma samples at 200-1000 fold. Due to individual differences, please estimate the concentration range of the sample in advance, and conduct a preliminary test to determine the appropriate dilution ratio of the sample.

If your test sample needs dilution, please refer to the dilution method as follows:
For 100 fold dilution: One-step dilution. Add 5 μL sample to 495 μL sample diluent to yield 100 fold dilution.
For 1000 fold dilution: Two-step dilution. Add 5 μL sample to 95 μL sample diluent to yield 20 fold dilution, then add 5 μL 20 fold diluted sample to 245 μL sample diluent, after this, the neat sample has been diluted at 1000 fold successfully.
For 100000 fold dilution: Three-step dilution. Add 5 μL sample to 195 μL sample diluent to yield 40 fold dilution, then add 5 μL 40 fold diluted sample to 245 μL sample diluent to yield 50 fold dilution, and finally add 5 μL 2000 fold diluted sample to 245 μL sample diluent, after this, the neat sample has been diluted at 100000 fold successfully.

Reagent preparation

1. Bring all reagents to room temperature (18-25℃) before use. If the kit will not be used up in one assay, please only take out the necessary strips and reagents for present experiment, and store the remaining strips and reagents at required condition.
2. **Wash Buffer:** Dilute 30 mL of Concentrated Wash Buffer with 720 mL of deionized or distilled water to prepare 750 mL of Wash Buffer. Note: if crystals have formed in the concentrate, warm it in a 40℃ water bath and mix it gently until the crystals have completely dissolved. For same day use only.
3. **Standard working solution:** Centrifuge the standard at 10,000×g for 1 min. Add 1 mL of Reference Standard &Sample Diluent, let it stand for 10 min and invert it gently several times. After it dissolves fully, mix it thoroughly with a pipette. This reconstitution produces a working solution of 25 ng/mL (or add 1 mL of Reference Standard &Sample Diluent, let it stand for 1-2 min and then mix it thoroughly with a vortex meter of low speed. Bubbles generated during vortex could be removed by centrifuging at a relatively low speed). Then make serial dilutions as needed. The recommended dilution gradient is as follows: 25, 12.5, 6.25, 3.13, 1.56, 0.78, 0.39, 0 ng/mL.
   Dilution method: Take 7 EP tubes, add 500 μL of Reference Standard & Sample Diluent to each tube. Pipette 500 μL of the 25 ng/mL working solution to the first tube and mix up to produce a 12.5 ng/mL working solution. Pipette 500 μL of the solution from the former tube into the latter one according to this step. The illustration on the next page is for reference. Note: the last tube is regarded as a blank. Don't pipette

solution into it from the former tube. Gradient diluted standard working solution should be prepared just before use.

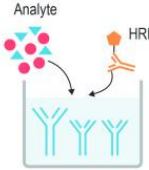


4. **HRP Conjugate working solution:** HRP Conjugate is HRP conjugated antibody. Calculate the required amount before the experiment (50 μL/well). In preparation, slightly more than calculated should be prepared. Centrifuge the **Concentrated HRP Conjugate** at 800×g for 1 min, then dilute the 100×**Concentrated HRP Conjugate** to 1× working solution with **HRP Conjugate Diluent**(Concentrated HRP Conjugate: HRP Conjugate Diluent= 1: 99). The working solution should be prepared just before use.

Assay procedure

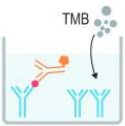
1. Determine wells for **diluted standard**, **blank** and **sample**. Add 50 μL each dilution of standard, blank and sample into the appropriate wells (It is recommended that all samples and standards be assayed in duplicate. It is recommended to determine the dilution ratio of samples through preliminary experiments or technical support recommendations). Immediately add 50 μL of **HRP Conjugate working solution** to each well. Cover the plate with a new sealer. Incubate for 60 min at 37℃. Note: solutions should be added to the bottom of the micro ELISA plate well, avoid touching the inside wall and causing foaming as much as possible.
2. Decant the solution from each well，add 350 μL of **wash buffer** to each well. Soak for 1 min and aspirate or decant the solution from each well and pat it dry against clean absorbent paper. Repeat this wash step 5 times. Note: a microplate washer can be used in this step and other wash steps. Make the tested strips in use immediately after the wash step. Do not allow wells to be dry.
3. Add 90 μL of **Substrate Reagent** to each well. Cover the plate with a new sealer. Incubate for about 15 min at 37℃. Protect the plate from light. Note: the reaction time can be shortened or extended according to the actual color change, but not more than 30 min. Preheat the Microplate Reader for about 15 min before OD measurement.
4. Add 50 μL of **Stop Solution** to each well. Note: adding the stop solution should be done in the same order as the substrate solution.
5. Determine the optical density (OD value) of each well at once with a micro-plate reader set to 450 nm.

Assay Procedure Summary



1. Add 50μL standard or sample to the wells, immediately add 50μL HRP Linked Detection Ab working solution to each well. Incubate for 60 min at 37°C

2. Aspirate and wash the plate for 5 times



3. Add 90 μL of Substrate Reagent. Incubate for about 15 min at 37°C

4. Add 50μL Stop Solution

5. Read the plate at 450nm immediately. Calculation of the results

Calculation of results

Average the duplicate readings for each standard and samples, then subtract the average zero standard optical density. Plot a four parameter logistic curve on log-log axis, with standard concentration on the x-axis and OD values on the y-axis.

If the OD of the sample surpasses the upper limit of the standard curve, you should re-test it with an appropriate dilution. The actual concentration is the calculated concentration multiplied by the dilution factor.

Technical resources

More detailed ELISA experiment guidelines and routine problem analysis can be obtained through wechat QR code at the lower left.

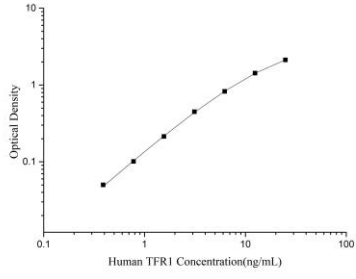
If you have any technical problems, please feel free to contact our technical support (it is recommended to take pictures and save the experimental data in time. Keep the used plate and remaining reagents).

Guidelines for ELISA

Wechat of technical support

Typical data

As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test. Typical standard curve and data is provided below for reference only.

ng/mL	OD	Corrected OD	Standard Curve
25	2.196	2.120	
12.5	1.498	1.422	
6.25	0.900	0.824	
3.13	0.523	0.447	
1.56	0.289	0.213	
0.78	0.177	0.101	
0.39	0.126	0.050	
0	0.076	-	

Sample value

Sample type	Reference range of Human TFR1 (ng/mL)
Serum(n=10)	400-2000
Plasma(EDTA)(n=10)	500-3000

Note: The above samples were from normal healthy individuals during non-menstrual period or pregnancy.

Performance

■ Precision

Intra-assay Precision (Precision within an assay): 3 samples with low, mid range and high level Human TFR1 were tested 20 times on one plate, respectively.

Inter-assay Precision (Precision between assays): 3 samples with low, mid range and high level Human TFR1 were tested on 3 different plates, 20 replicates in each plate, respectively.

	Intra-assay Precision			Inter-assay Precision		
Sample	1	2	3	1	2	3
n	20	20	20	20	20	20
Mean(ng/mL)	1.26	2.27	8.84	1.25	2.42	8.91
Standard deviation	0.07	0.13	0.38	0.09	0.15	0.54
CV (%)	5.61	5.71	4.35	7.12	6.1	6.01

■ Recovery

The recovery of Human TFR1 spiked at three different levels in samples throughout the range of the assay was evaluated in various matrices.

Sample Type	Range (%)	Average Recovery (%)
Serum (n=8)	91-104	96
EDTA plasma (n=8)	93-108	101

■ Linearity

Samples were spiked with high concentrations of Human TFR1 and diluted with Reference Standard & Sample Diluent to produce samples with values within the range of the assay.

		Serum (n=5)	EDTA plasma (n=5)
1:2	Range (%)	95-111	95-106
	Average (%)	102	100
1:4	Range (%)	96-113	89-103
	Average (%)	104	95
1:8	Range (%)	95-107	95-109
	Average (%)	101	102
1:16	Range (%)	95-111	89-102
	Average (%)	101	96

Declaration

1. Limited by current conditions and scientific technology, we can't conduct comprehensive identification and analysis on all the raw material provided. So there might be some qualitative and technical risks for users using the kit.
2. This assay is designed to eliminate interference by factors present in biological samples. Until all factors have been tested in the ELISA immunoassay, the possibility of interference cannot be excluded.
3. The final experimental results will be closely related to the validity of products, operational skills of the operators, the experimental environments and so on. We are only responsible for the kit itself, but not for the samples consumed during the assay. The users should calculate the possible amount of the samples used in the whole test. Please reserve sufficient samples in advance.
4. To get the best results, please only use the reagents supplied by the manufacturer and strictly comply with the instructions.
5. Incorrect results may occur because of incorrect operations during the reagents preparation and loading, as well as incorrect parameter settings of the Micro-plate reader. Please read the instructions carefully and adjust the instrument prior to the experiment.
6. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
7. Every kit has strictly passed QC test. However, results from end users might be inconsistent with our data due to some variables such as transportation conditions, different lab equipment, and so on. Intra-assay variance among kits from different batches might arise from the above reasons too.
8. Kits from different manufacturers or other methods for testing the same analyte could bring out inconsistent results, since we haven't compared our products with those from other manufacturers.
9. The kit is designed for research use only, we will not be responsible for any issues if the kit is applied in clinical diagnosis or any other related procedures.