（FOR RESEARCH USE ONLY. DO NOT USE IT IN CLINICAL DIAGNOSTICS !）

Catalog No: CQH001

Product size: 96T/48T/24T/96T*5

Elabscience® CellaQuant™ Human IL-6 (Interleukin 6) ELISA Kit

This manual must be read attentively and completely before using this product.

If you have any problems, please contact our Technical Service Center for help (info in the header of each page).

Tel: 1-832-243-6086
Fax: 1-832-243-6017
Email: techsupport@elabscience.com
Website: www.elabscience.com

Please refer to specific expiry date from label outside of box.

Please kindly provide us with the lot number (on the outside of the box) of the kit for more efficient service.

Intended use

This ELISA kit applies to the in vitro quantitative determination of Human IL-6 concentrations in cell supernatant, cell lysate and other biological fluids.

Character

Item	
Sensitivity	2.81 pg/mL
Detection Range	4.69-300 pg/mL
Specificity	This kit recognizes Human IL-6 in samples. No significant cross-reactivity or interference between Human IL-6 and analogues was observed
Repeatability	Coefficient of variation is < 10%

Test principle

This ELISA kit uses the Sandwich-ELISA principle. The micro ELISA plate provided in this kit has been pre-coated with an antibody specific to Human IL-6. Samples (or Standards &Quality Control Material) are added to the micro ELISA plate wells and combined with the specific antibody. Then a biotinylated detection antibody specific for Human IL-6 and Avidin-Horseradish Peroxidase (HRP) conjugate are added successively to each micro plate well and incubated. Free components are washed away. The substrate solution is added to each well. Only those wells that contain Human IL-6, biotinylated detection antibody and Avidin-HRP conjugate will appear blue in color. The enzyme-substrate reaction is terminated by the addition of stop solution and the color turns yellow. The optical density (OD) is measured spectrophotometrically at a wavelength of 450±2 nm. The OD value is proportional to the concentration of Human IL-6. You can calculate the concentration of Human IL-6 in the samples by comparing the OD of the samples to the standard curve.

Kit components & Storage

An unopened kit can be stored at 2-8℃ for 12 months. After opening, store the items separately according to the following conditions.

Item	Specifications	Storage
Micro ELISA Plate (Dismountable)	96T: 8 wells ×12 strips 48T: 8 wells ×6 strips 24T: 8 wells ×3 strips 96T*5: 5 plates, 96T	-20℃, up to expiry date (12 months)
Reference Standard	96T: 2 vials 48T/24T: 1 vial 96T*5: 10 vials	
Concentrated Biotinylated Detection Ab(100×)	96T: 1 vial, 120 µL 48T/24T: 1 vial, 60 µL 96T*5: 5 vials, 120 µL	
Concentrated HRP Conjugate (100×)	96T: 1 vial, 120 µL 48T/24T: 1 vial, 60 µL 96T*5: 5 vials, 120 µL	
Quality Control Material	96T: 2 vials 48T/24T: 1 vial 96T*5: 10 vials	
Reference Standard & Quality Control Material &Sample Diluent	96T/48T/24T:2 vials, 20 mL 96T*5: 10 vials, 20 mL	2-8℃, up to expiry date (12 months)
Biotinylated Detection Ab Diluent	96T/48T/24T: 1 vial, 14 mL 96T*5: 5 vials, 14 mL	
HRP Conjugate Diluent	96T/48T/24T: 1 vial, 14 mL 96T*5: 5 vials, 14 mL	
Concentrated Wash Buffer(25×)	96T/48T/24T: 1 vial, 30 mL 96T*5: 5 vials, 30 mL	
Substrate Reagent	96T/48T/24T: 1 vial, 10 mL 96T*5: 5 vials, 10 mL	
Stop Solution	96T/48T/24T: 1 vial, 10 mL 96T*5: 5 vials, 10 mL	
Plate Sealer	96T/48T/24T: 5 pieces 96T*5: 25 pieces	
Product Description	1 copy	

Note: Concentrated HRP Conjugate(100×) and **Substrate Reagent** should be stored away from light.

All reagent bottle caps must be tightened to prevent evaporation and microbial pollution. The volume of reagents in partial shipments is a little more than the volume marked on the label, please use accurate

measuring equipment instead of directly pouring into the vial(s).

Other supplies required

Microplate reader with 450 nm wavelength filter
High-precision transfer pipette, EP tubes and disposable pipette tips
Incubator capable of maintaining 37℃
Deionized or distilled water
Absorbent paper
Loading slot

Sample collection

Cell culture supernatant or other biological fluids: Centrifuge samples for 20 min at 1000×g at 2-8℃. Collect the supernatant to carry out the assay.
Cell lysates: For adherent cells, gently wash the cells with moderate amount of pre-cooled PBS and dissociate the cells using trypsin. Collect the cell suspension into a centrifuge tube and centrifuge for 5 min at 1000×g. Discard the medium and wash the cells 3 times with pre-cooled PBS. For each 1×10^6 cells, add 150-250 µL of pre-cooled PBS to keep the cells suspended. Repeat the freeze-thaw process several times or use an ultrasonic cell disrupter until the cells are fully lysed. Centrifuge for 10 min at 1500×g at 2-8℃. Remove the cell fragments, collect the supernatant to carry out the assay.
Recommended reagents for sample preparation: PMSF Protease Inhibitor (Cat No. E-EL-SR002), 0.25% Trypsin Solution (Cat No. E-EL-SR001).

Note

■ Note for kit

1) For research use only. Not for use in diagnostic procedures.
2) Please wear lab coats, eye protection and latex gloves for protection. Please perform the experiment following the national security protocols of biological laboratories, especially when detecting blood samples or other bodily fluids.
3) A freshly opened ELISA plate may appear a water-like substance, which is normal and will not have any impact on the experimental results. Return the unused wells to the foil pouch and store according to the conditions suggested in the above table.
4) Do not reuse the reconstituted standard, Quality Control Material, biotinylated detection Ab working solution, HRP conjugate working solution. The unspent undiluted concentrated biotinylated detection Ab (100×) and other stock solutions should be stored according to the storage conditions in the above table.
5) The microplate reader should be able to be installed with a filter that can detect the wave length at 450±2 nm. The optical density should be within 0-3.5. Follow the Instructions of the Microplate Reader for set-up and preheat it for 15 min before OD measurement.
6) **Do not mix or substitute reagents with those from other lots or sources.**
7) Change pipette tips in between adding of each standard level, between sample adding and between reagent adding. Also, use separate reservoirs for each reagent.
8) The kit should not be used beyond the expiration date on the kit label.

■ Note for sample

1) Tubes for blood collection should be disposable and be non-endotoxin. Samples with high hemolysis or much lipid are not suitable for ELISA assay.
2) Samples should be assayed within 7 days when stored at 2-8℃, otherwise samples must be divided up and stored at -20℃ (≤1 month) or -80℃ (≤3 months). Avoid repeated freeze-thaw cycles. Prior to assay, the frozen samples should be slowly thawed and centrifuged to remove precipitates.
3) Please predict the concentration before assaying. If the sample concentration is not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments.
4) If the sample type is not included in the manual, a preliminary experiment is suggested to verify the validity.
5) If a lysis buffer is used to prepare tissue homogenates or cell lysates,

there is a possibility of causing a deviation due to the introduced chemical substance.

6) Some recombinant protein may not be detected due to a mismatching with the coated antibody or detection antibody.

Dilution method

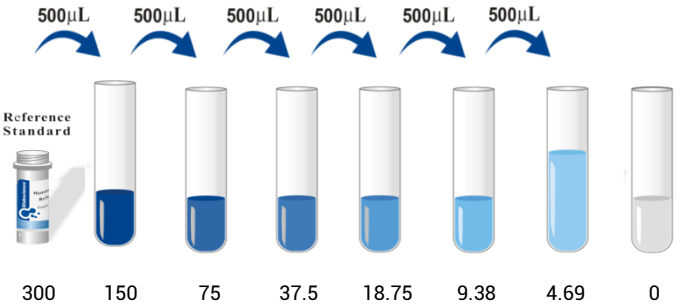
Please predict the concentration range of samples in advance, and determine the dilution ratio through preliminary experiments or technical support recommendations.

If your test sample needs dilution, please refer to the dilution method as follows:
For 100 fold dilution: One-step dilution. Add 5 µL sample to 495 µL sample diluent to yield 100 fold dilution.
For 1000 fold dilution: Two-step dilution. Add 5 µL sample to 95 µL sample diluent to yield 20 fold dilution, then add 5 µL 20 fold diluted sample to 245 µL sample diluent, after this, the neat sample has been diluted at 1000 fold successfully.
For 100000 fold dilution: Three-step dilution. Add 5 µL sample to 195 µL sample diluent to yield 40 fold dilution, then add 5 µL 40 fold diluted sample to 245 µL sample diluent to yield 50 fold dilution, and finally add 5 µL 2000 fold diluted sample to 245 µL sample diluent, after this, the neat sample has been diluted at 100000 fold successfully.

Reagent preparation

1. Bring all reagents to room temperature (18-25℃) before use. If the kit will not be used up in one assay, please only take out the necessary strips and reagents for present experiment, and store the remaining strips and reagents at required condition.
2. **Wash Buffer:** Dilute 30 mL of Concentrated Wash Buffer with 720 mL of deionized or distilled water to prepare 750 mL of Wash Buffer. Note: if crystals have formed in the concentrate, warm it in a 40℃ water bath and mix it gently until the crystals have completely dissolved. For same day use only.

3. Standard working solution: Centrifuge the standard at 10,000×g for 1 min. Add 1 mL of Reference **Standard & Quality Control Material& Sample Diluent,** let it stand for 10 min and invert it gently several times. After it dissolves fully, mix it thoroughly with a pipette. This reconstitution produces a working solution of 300 pg/mL (or add 1 mL of **Reference Standard & Quality Control Material& Sample Diluent**, let it stand for 1-2 min and then mix it thoroughly with a vortex meter of low speed. Bubbles generated during vortex could be removed by centrifuging at a relatively low speed). Then make serial dilutions as needed. The recommended dilution gradient is as follows: 300, 150, 75, 37.5, 18.75, 9.38, 4.69, 0 pg/mL.
   Dilution method: Take 7 EP tubes, add 500 µL of **Reference Standard & Quality Control Material&Sample Diluent** to each tube. Pipette 500 µL of the 300 pg/mL working solution to the first tube and mix up to produce a 150 pg/mL working solution. Pipette 500 µL of the solution from the former tube into the latter one according to this step. The illustration on the next page is for reference. Note: the last tube is regarded as a blank. Don't pipette solution into it from the former tube.
   The working solution of the standard substance at 300 pg/mL after reconstitution should be aliquoted and stored at -20°C.It should be used up within half a month and repeated freeze-thaw should be avoided.
   Gradient diluted standard working solution should be prepared just before use.



4. **Quality Control Standard working solution**: Centrifuge the Quality Control Standard at 10,000×g for 1 min. Add 1000 μL of Reference Standard & Quality Control Material& Sample Diluent, let it stand for 10 min and invert it gently several times. After it dissolves fully, mix it thoroughly with a pipette. This reconstitution produces a working solution of (or add 1000 μL of Reference Standard & Quality Control Material& Sample Diluent, let it stand for 1-2 min and then mix it thoroughly with a vortex meter of low speed. Bubbles generated during vortex could be removed by centrifuging at a relatively low speed).
   Quality Control Material working solution should be prepared just before use.
5. **Biotinylated Detection Ab working solution**: Calculate the required amount before the experiment (100 μL/well). In preparation, slightly more than calculated should be prepared. Centrifuge the Concentrated Biotinylated Detection Ab at 800×g for 1 min, then dilute the 100× Concentrated Biotinylated Detection Ab to 1× working solution with Biotinylated Detection Ab Diluent (Concentrated Biotinylated Detection Ab: Biotinylated Detection Ab Diluent= 1: 99). The working solution should be prepared just before use.
6. **HRP Conjugate working solution:** HRP Conjugate is HRP conjugated avidin. Calculate the required amount before the experiment (100 μL/well). In preparation, slightly more than calculated should be prepared. Centrifuge the Concentrated HRP Conjugate at 800×g for 1 min, then dilute the 100× Concentrated HRP Conjugate to 1× working solution with HRP Conjugate Diluent (Concentrated HRP Conjugate: HRP Conjugate Diluent= 1: 99). The working solution should be prepared just before use.

Assay procedure

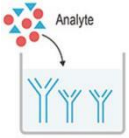
1. Determine wells for **diluted standard**, **quality control material, blank** and **sample**. Add 100 µL each dilution of standard, quality control material, blank and sample into the appropriate wells (It is recommended that all samples and standards be assayed in duplicate. It is recommended to determine the dilution ratio of samples through preliminary experiments or technical support recommendations). Cover the plate with the sealer provided in the kit. Incubate for 90 min at 37℃. .Note: solutions should be added to the bottom of the micro ELISA plate well, avoid touching the inside wall and causing foaming as much as possible.
2. Decant the liquid from each well, do not wash. Immediately add 100 µL of **Biotinylated Detection Ab working solution** to each well. Cover the plate with a new sealer. Incubate for 1 hour at 37°C.
3. Decant the solution from each well， add 350 µL of **wash buffer** to each well. Soak for 1 min and aspirate or decant the solution from each well and pat it dry against clean absorbent paper. Repeat this wash step 3 times. Note: a microplate washer can be used in this step and other wash steps. Make the tested strips in use immediately after the wash step. Do not allow wells to be dry.
4. Add 100 µL of **HRP Conjugate working solution** to each well. Cover the plate with a new sealer. Incubate for 30 min at 37°C.
5. Decant the solution from each well, repeat the wash process for 5 times as conducted in step 3.
6. Add 90 µL of **Substrate Reagent** to each well. Cover the plate with a new sealer. Incubate for about 15 min at 37°C. Protect the plate from light. Note: the reaction time can be shortened or extended according to the actual color change, but not more than 30 min. Preheat the Microplate Reader for about 15 min before OD measurement.
7. Add 50 µL of **Stop Solution** to each well. Note: adding the stop solution should be done in the same order as the substrate solution.
8. Determine the optical density (OD value) of each well at once with a micro-plate reader set to 450 nm.

Calculation of results

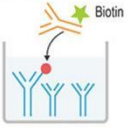
Average the duplicate readings for each standard and samples, then subtract the average zero standard optical density. Plot a four parameter logistic curve on log-log axis, with standard concentration on the x-axis and OD values on the y-axis.

If the OD of the sample surpasses the upper limit of the standard curve, you should re-test it with an appropriate dilution. The actual concentration is the calculated concentration multiplied by the dilution factor.

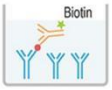
Assay Procedure Summary



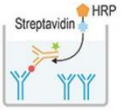
1. Add 100μL standard, quality control material or sample to the wells. Incubate for 90 min at 37℃



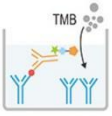
2. Discard the liquid, immediately add 100μL Biotinylated Detection Ab working solution to each well. Incubate for 60 min at 37℃



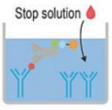
3. Aspirate and wash the plate for 3 times



4. Add 100μL HRP conjugate working solution. Incubate for 30 min at 37℃. Aspirate and wash the plate for 5 times



5. Add 90μL Substrate Reagent. Incubate for 15 min at 37℃

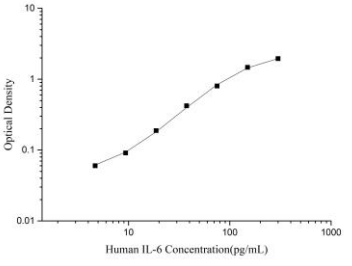


6. Add 50μL Stop Solution

7. Read the plate at 450nm immediately. Calculation of the results

Typical data

As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test. Typical standard curve and data is provided below for reference only.

pg/mL	OD	Corrected OD	Standard Curve
300	2.033	1.946	
150	1.553	1.466	
75	0.889	0.802	
37.5	0.508	0.421	
18.75	0.275	0.188	
9.38	0.178	0.091	
4.69	0.147	0.060	
0	0.087	-	

Performance

■ Precision

Intra-assay Precision (Precision within an assay): 3 samples with low, mid range and high level Human IL-6 were tested 20 times on one plate, respectively.

Inter-assay Precision (Precision between assays): 3 samples with low, mid range and high level Human IL-6 were tested on 3 different plates, 20 replicates in each plate, respectively.

	Intra-assay Precision			Inter-assay Precision		
Sample	1	2	3	1	2	3
n	20	20	20	20	20	20
Mean(pg/mL)	46.01	141.23	371.85	47.92	129.70	365.55
Standard deviation	2.35	8.50	17.96	3.02	9.34	27.23
CV (%)	5.11	6.02	4.83	6.31	7.20	7.45

■ Recovery

The recovery of Human IL-6 spiked at three different levels in samples throughout the range of the assay was evaluated in various matrices.

Sample Type	Range (%)	Average Recovery (%)
Human PBMC(n=6)	87-100	94
Human THP-1(n=6)	88-99	94
Human A549(n=6)	94-107	99

■ Linearity

Samples were spiked with high concentrations of Human IL-6 and diluted with Reference Standard & Sample Diluent to produce samples with values within the range of the assay.

		Human PBMC(n=5)	Human THP-1 (n=5)	Human A549 (n=5)
1:2	Range (%)	98-115	96-107	86-101
	Average (%)	105	102	93
1:4	Range (%)	101-116	92-107	87-99
	Average (%)	108	98	93
1:8	Range (%)	100-112	93-105	87-101
	Average (%)	106	99	93
1:16	Range (%)	97-115	93-104	89-101
	Average (%)	105	99	94

■ Sensitivity

The minimum detectable level of Human IL-6 is typically less than 2.81 pg/mL.

The MDD was determined by adding two standard deviations to the mean optical density (OD) of twenty replicate zero-standard wells and calculating the corresponding concentration

■ Description of Quality Control Material

The quality control material should be used directly after reconstitution (no dilution required) and tested as an unknown sample.

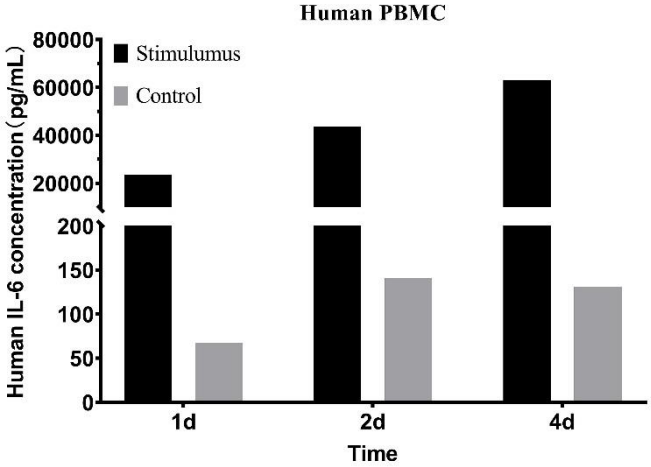
The acceptable range for the quality control material is M ± 3SD. However, due to potential variations in experimental techniques and methodologies, it is recommended that each laboratory establish its own acceptable range for the target values. Laboratories using other detection systems should also determine their own acceptable ranges, as results may vary across different detection systems.

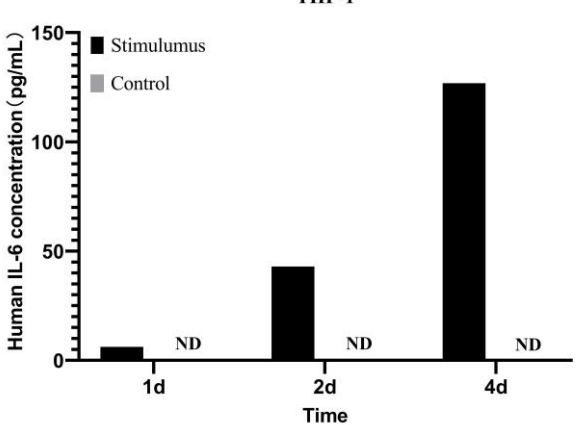
Target	Diluent Volume	M±3SD（pg/mL）
Human IL-6	1000 μL	62.72-76.74

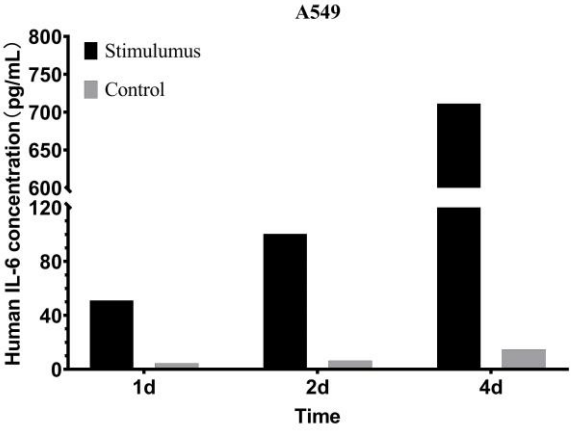
■ Cell Sample Values

1.Human PBMC cells ($1*10^6$cell/mL) were cultured in RPMI1640 supplemented with 10% fetal bovine serum. The culture medium also contained 1% Penicillin-Streptomycin Solution.
2.Human THP-1 cells ($5*10^5$cell/mL) were cultured in RPMI1640 supplemented with 10% fetal bovine serum. The culture medium also contained 1% Penicillin-Streptomycin Solution and 0.05 mM β-mercaptoethanol.
3. Human A549 cells (80% confluency) were cultured in Ham's F-12K supplemented with 10% fetal bovine serum. The culture medium also contained 1% Penicillin-Streptomycin Solution.
The cells were stimulated with different stimulants as listed in the table below, and the cell culture supernatant was collected to determine IL-6 content.

Cell Type	Stimulated Method	1 DAY (pg/mL)	2 DAY (pg/mL)	4 DAY (pg/mL)
Human PBMC	Control	67	141	131
	10 μg/mL PHA	23574	43562	63077
Human THP-1	Control	ND	ND	ND
	100 ng/mL PMA+500ng/mL LPS	6	42	126
Human A549	Control	4	6	14
	0.5μg/mL LPS	51	100	711


Human PBMC
Stimulumus
Control
Human IL-6 concentration（pg/mL）
80000
60000
40000
20000
200
150
100
50
0
1d
2d
4d
Time


THP-1
Stimulumus
Control
Human IL-6 concentration（pg/mL）
150
100
50
0
ND
ND
ND
1d
2d
4d
Time



- **Specificity**

  This ELISA method is capable of detecting both natural and recombinant Human IL-6 protein. To assess cross-reactivity with Human IL-6, the following factors were prepared at a concentration of 5 ng/mL using the dilution buffer (1×). No significant cross-reactivity or interference was observed.

Human	Others
IL-6R gp130	Mouse IL-6 Rat IL-6 Porcine IL-6

Declaration

1. Limited by current conditions and scientific technology, we can't conduct comprehensive identification and analysis on all the raw material provided. So there might be some qualitative and technical risks for users using the kit.
2. This assay is designed to eliminate interference by factors present in biological samples. Until all factors have been tested in the ELISA immunoassay, the possibility of interference cannot be excluded.
3. The final experimental results will be closely related to the validity of products, operational skills of the operators, the experimental environments and so on. We are only responsible for the kit itself, but not for the samples consumed during the assay. The users should calculate the possible amount of the samples used in the whole test. Please reserve sufficient samples in advance.
4. To get the best results, please only use the reagents supplied by the manufacturer and strictly comply with the instructions.
5. Incorrect results may occur because of incorrect operations during the reagents preparation and loading, as well as incorrect parameter settings of the Micro-plate reader. Please read the instructions carefully and adjust the instrument prior to the experiment.
6. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
7. Every kit has strictly passed QC test. However, results from end users might be inconsistent with our data due to some variables such as transportation conditions, different lab equipment, and so on. Intra-assay variance among kits from different batches might arise from the above reasons too.
8. Kits from different manufacturers or other methods for testing the same analyte could bring out inconsistent results, since we haven't compared our products with those from other manufacturers.
9. The kit is designed for research use only, we will not be responsible for any issues if the kit is applied in clinical diagnosis or any other relate